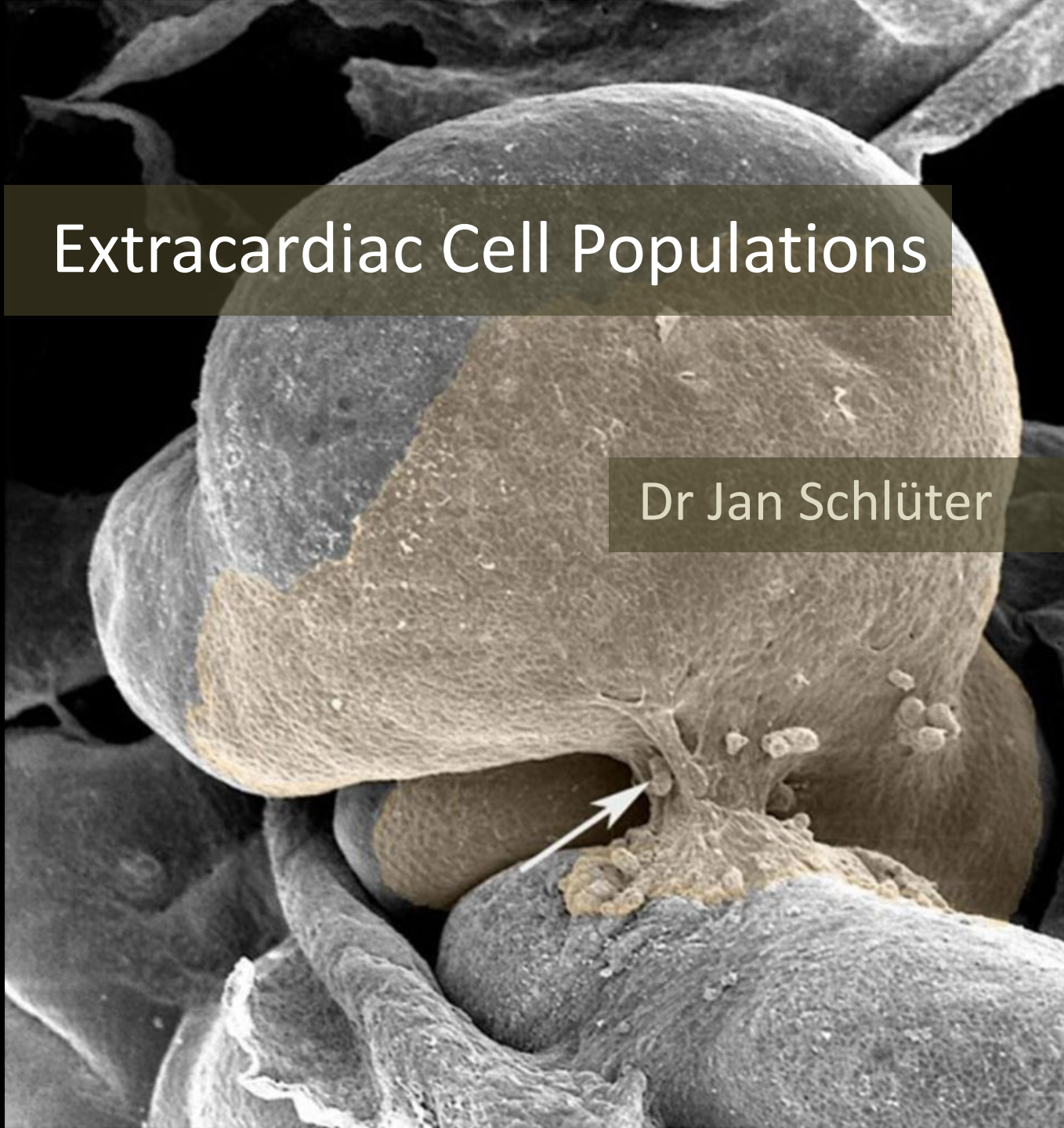


Extracardiac Cell Populations

Dr Jan Schlüter

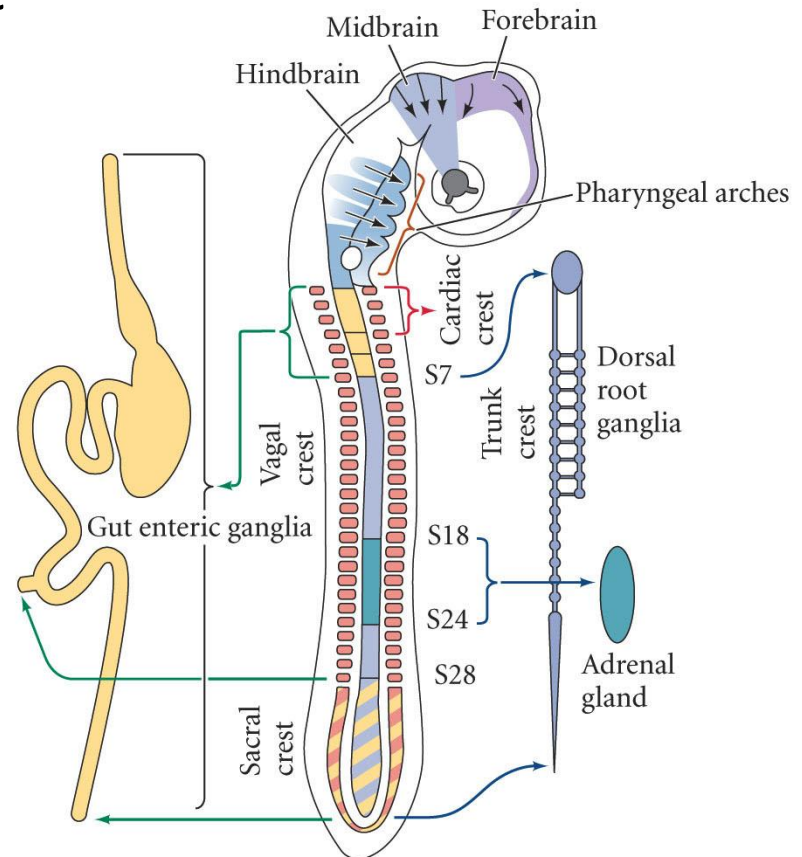


Extracardiac Cell Populations

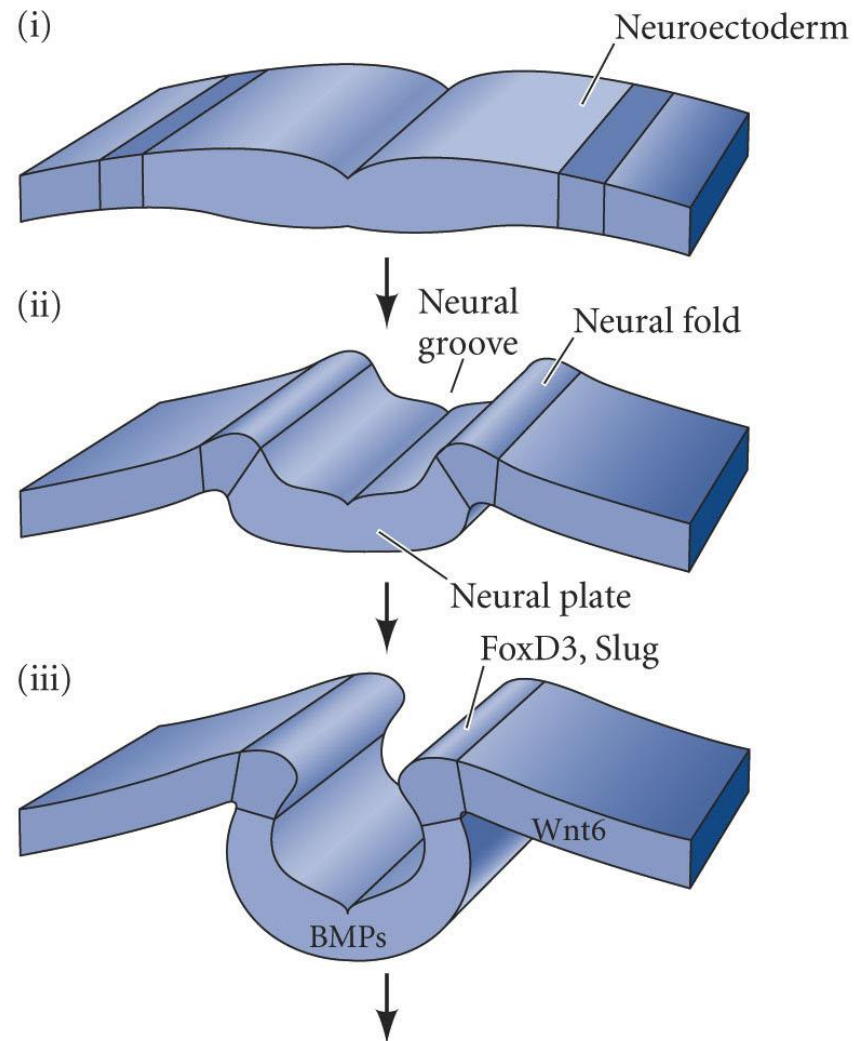
- I Neural Crest Cells
- II Proepicardial Cells

I Neural Crest Cells

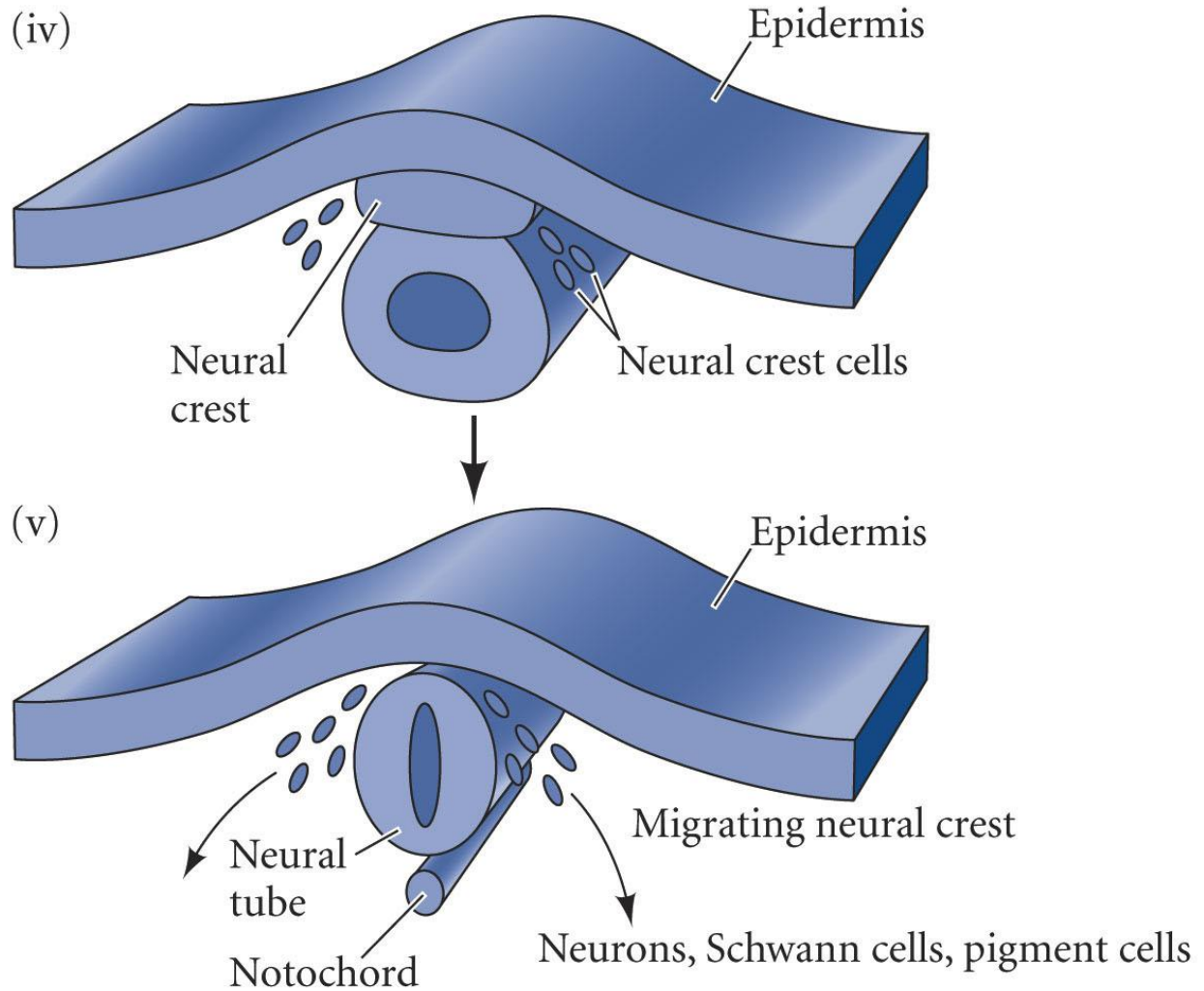
- Neural crest cells are multipotent cells which delaminate from the neural tube and migrate throughout the body
- Cranial (cephalic) neural crest
- Trunk neural crest
- Vagal and sacral neural crest
- Cardiac neural crest



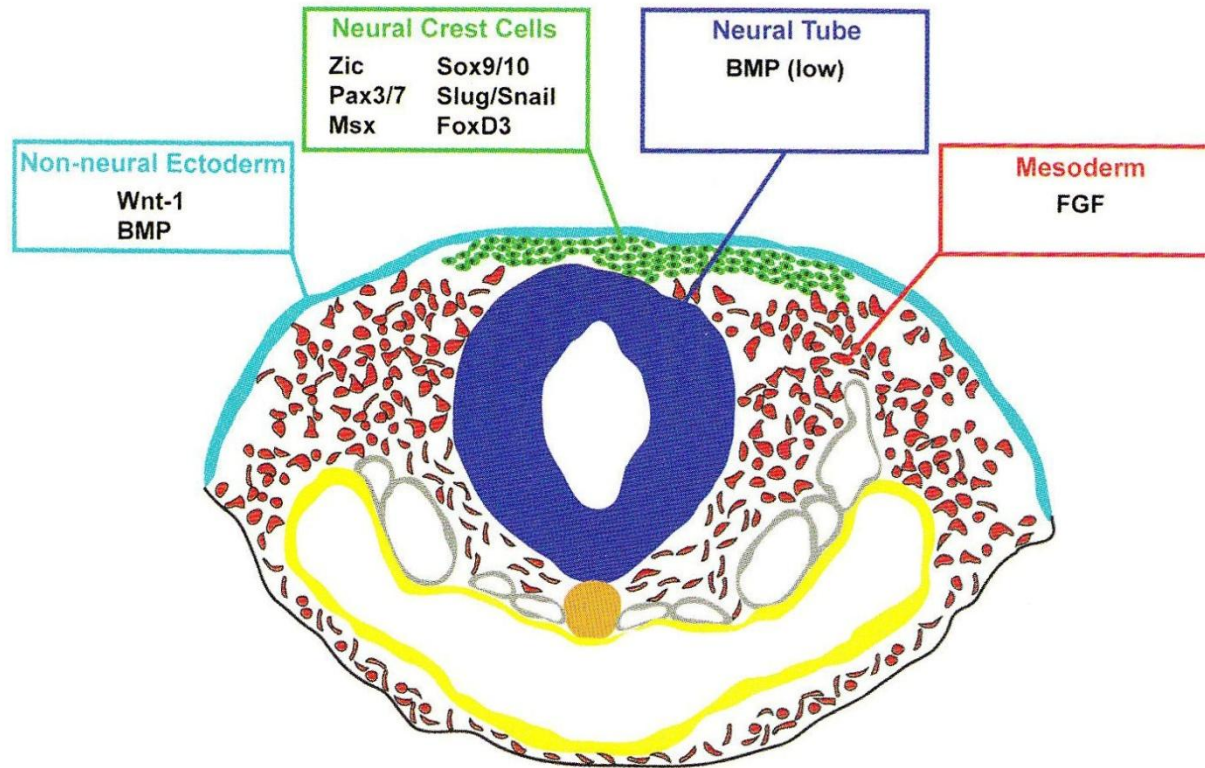
I Neural Crest Cells



I Neural Crest Cells



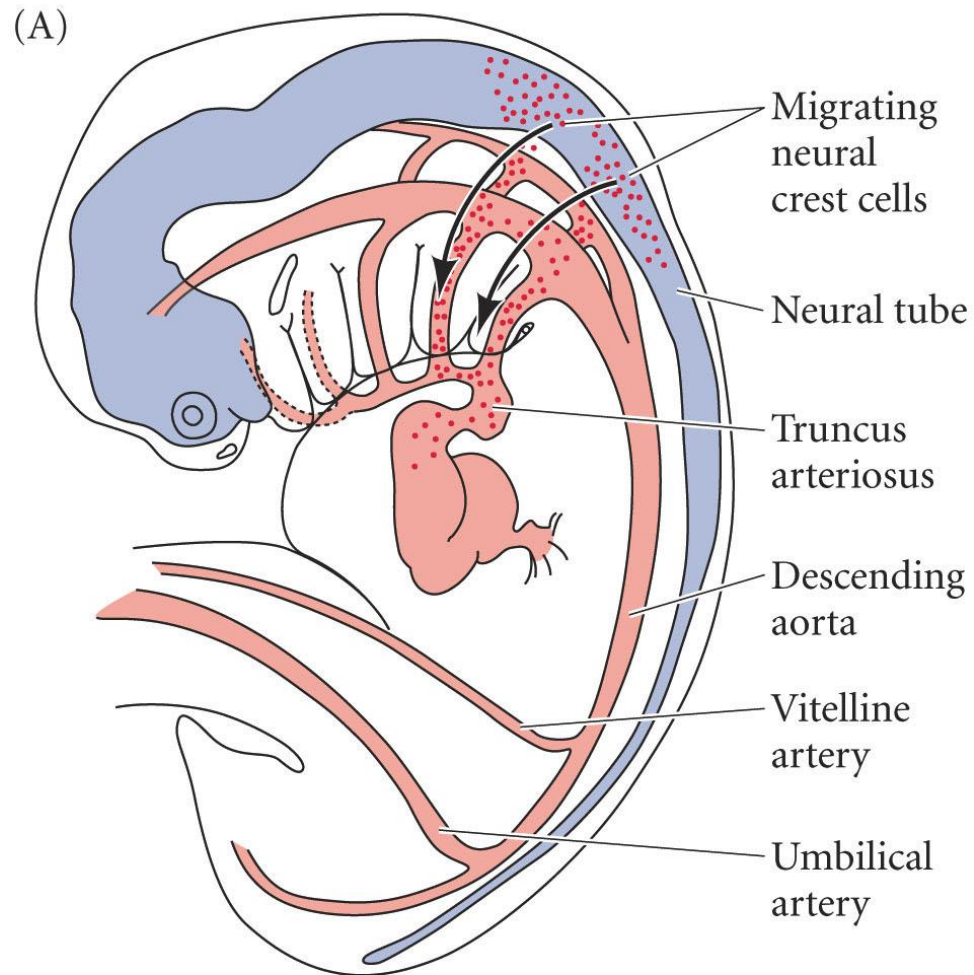
I Neural Crest Cells



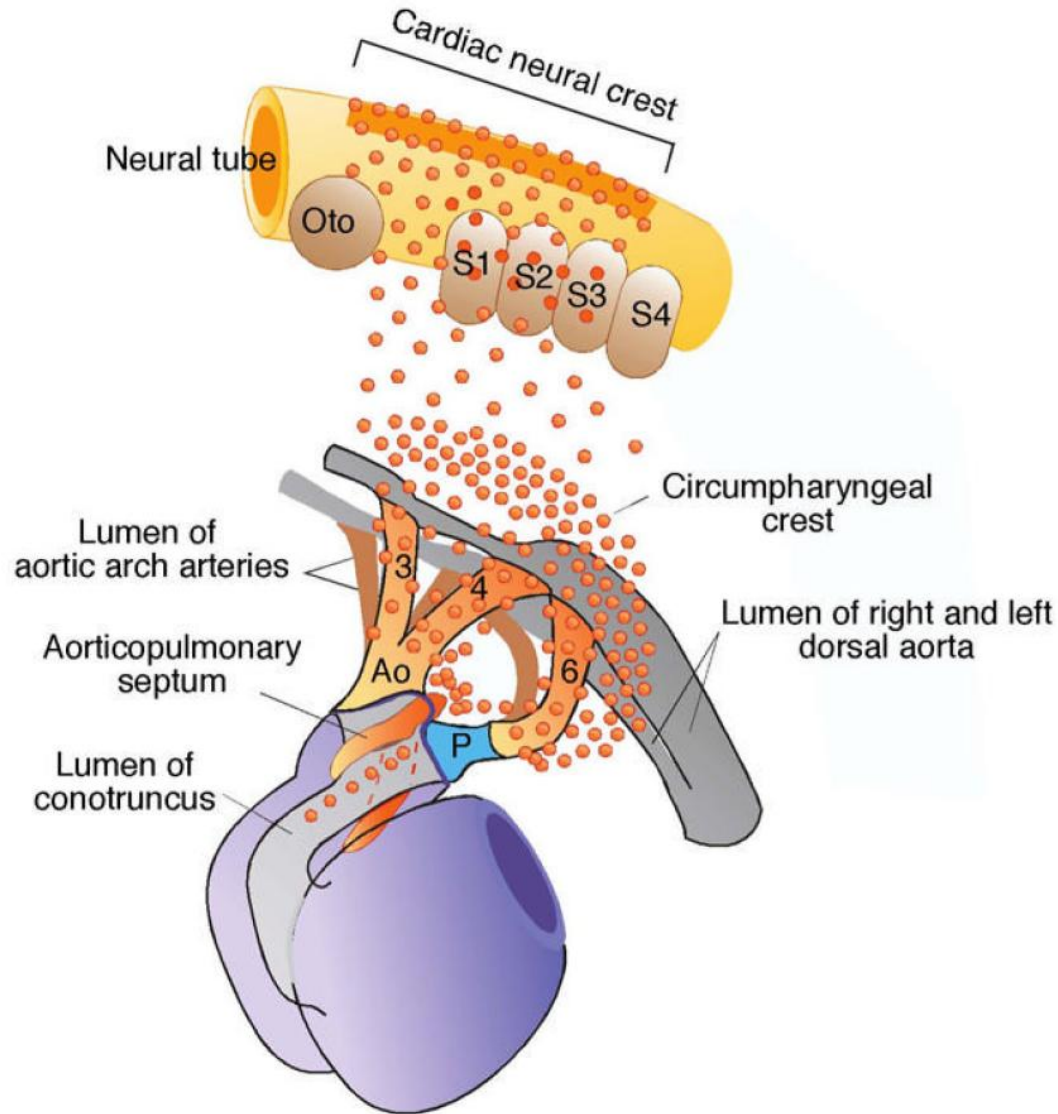
Cardiac neural crest cells

- Multipotent cardiac neural crest cells (CNCs) contribute to:
- Vascular smooth muscle
- Cushion mesenchyme
- Cardiac innervation

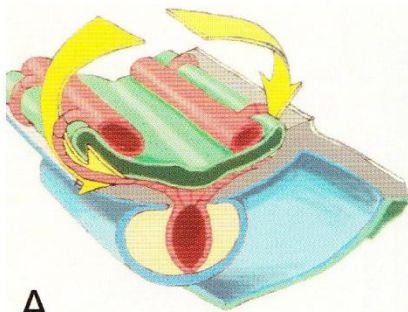
Cardiac neural crest cells



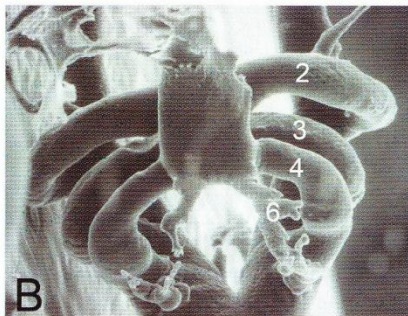
Cardiac neural crest cells



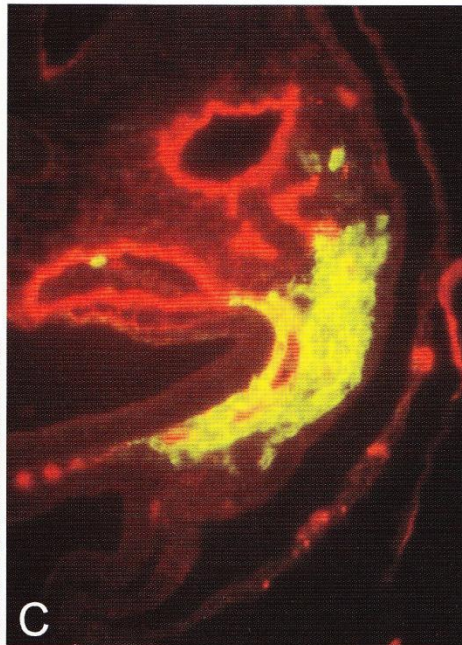
Cardiac neural crest cells



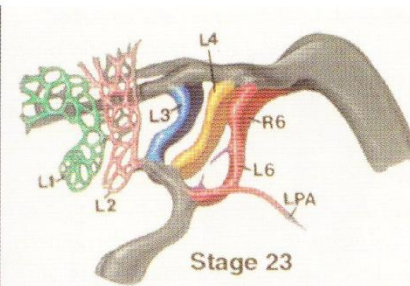
A



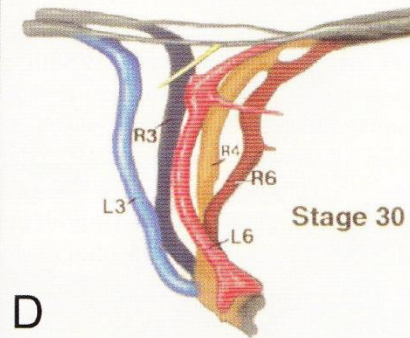
B



C



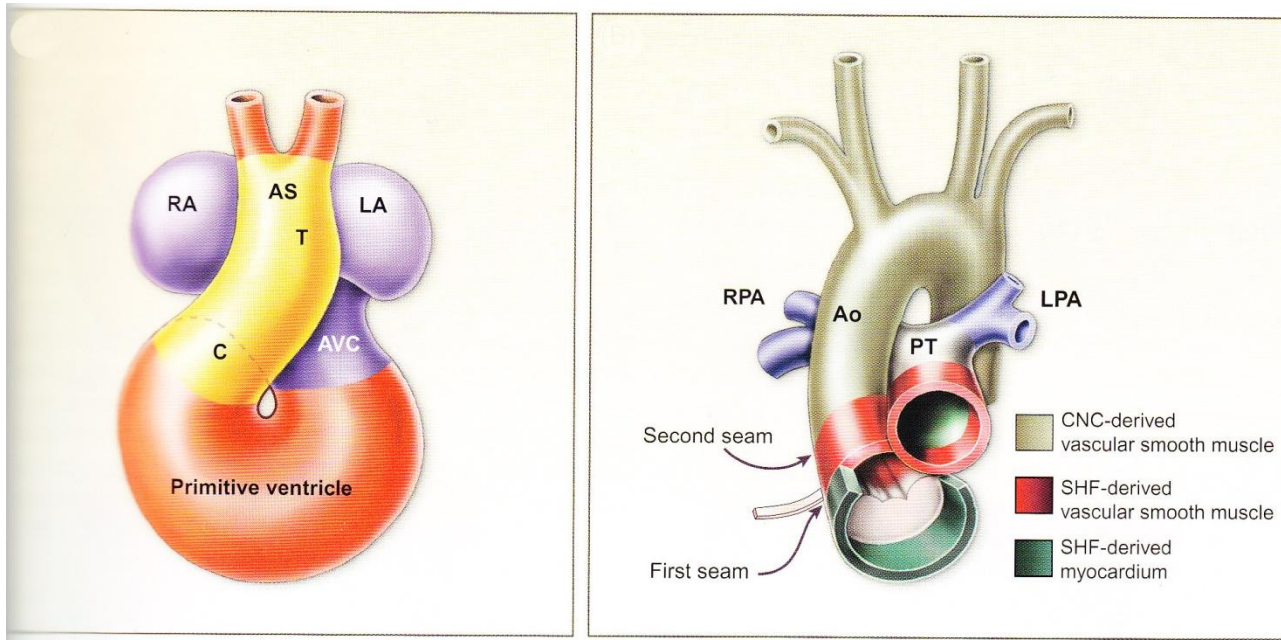
Stage 23



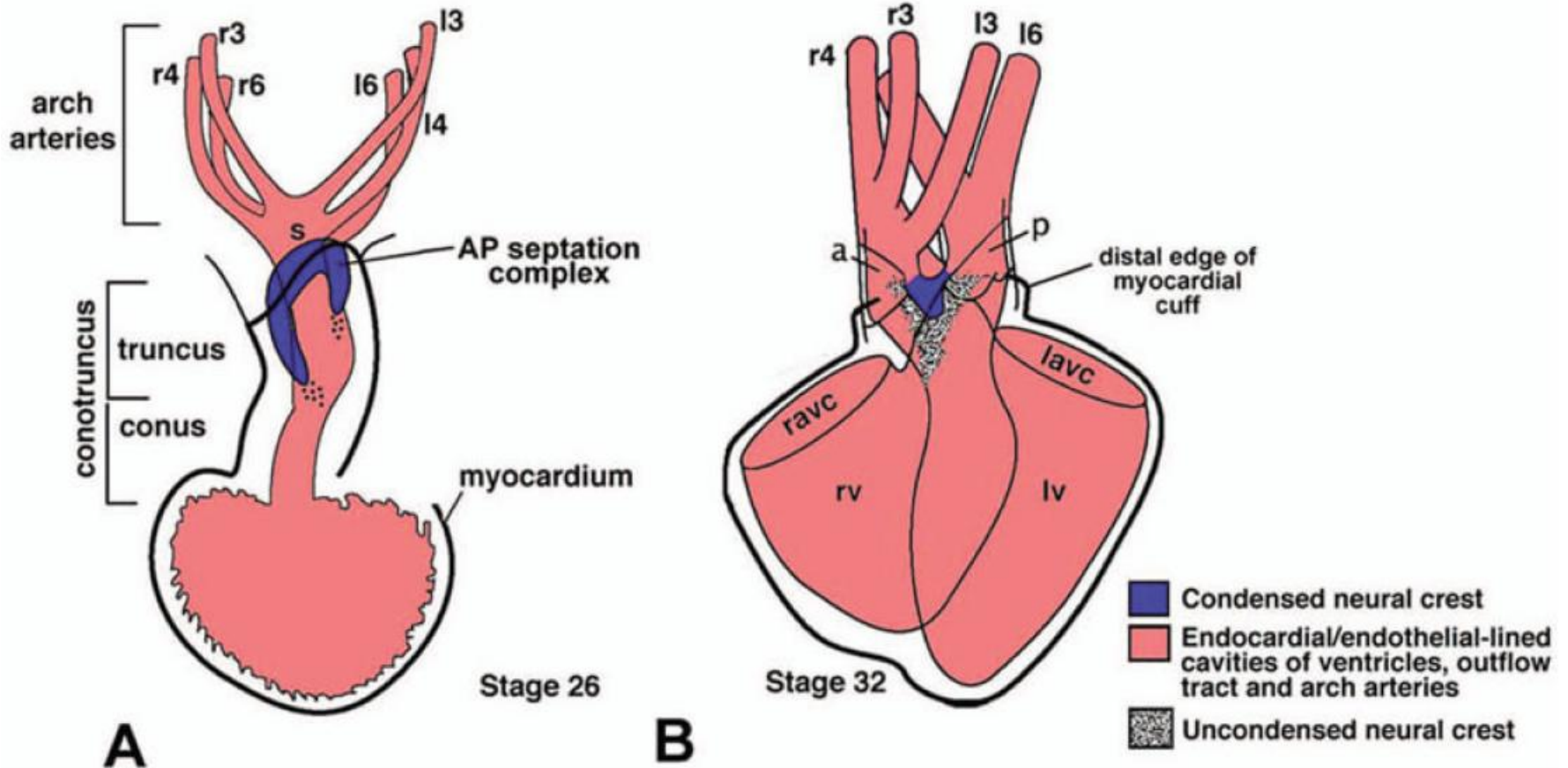
Stage 30

D

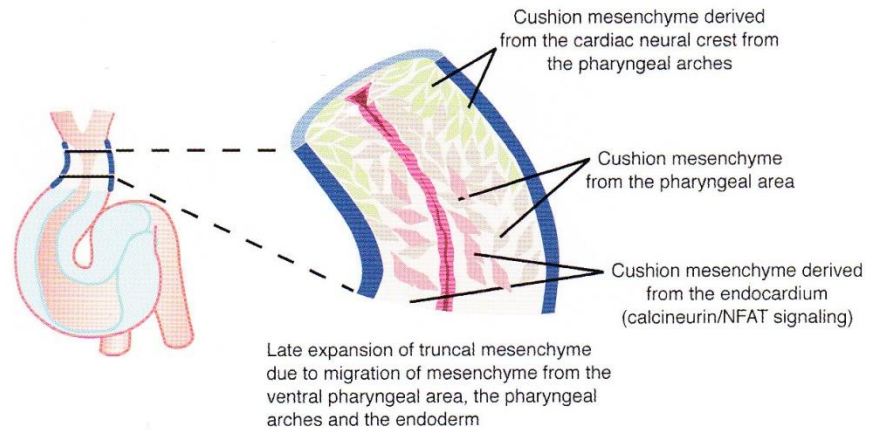
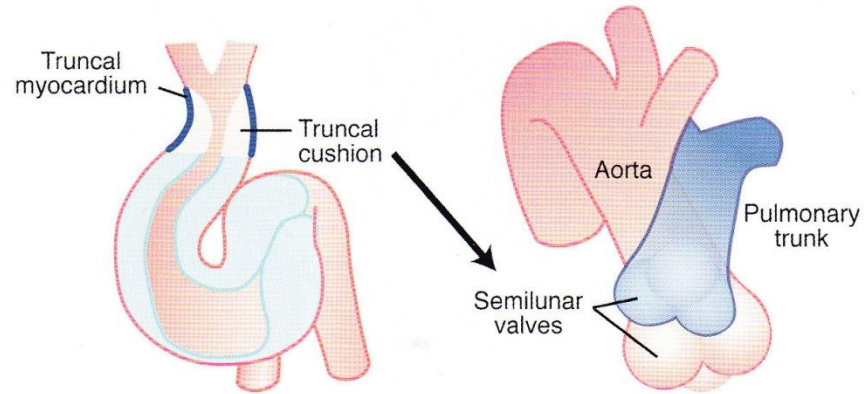
Cardiac neural crest cells



Cardiac neural crest cells

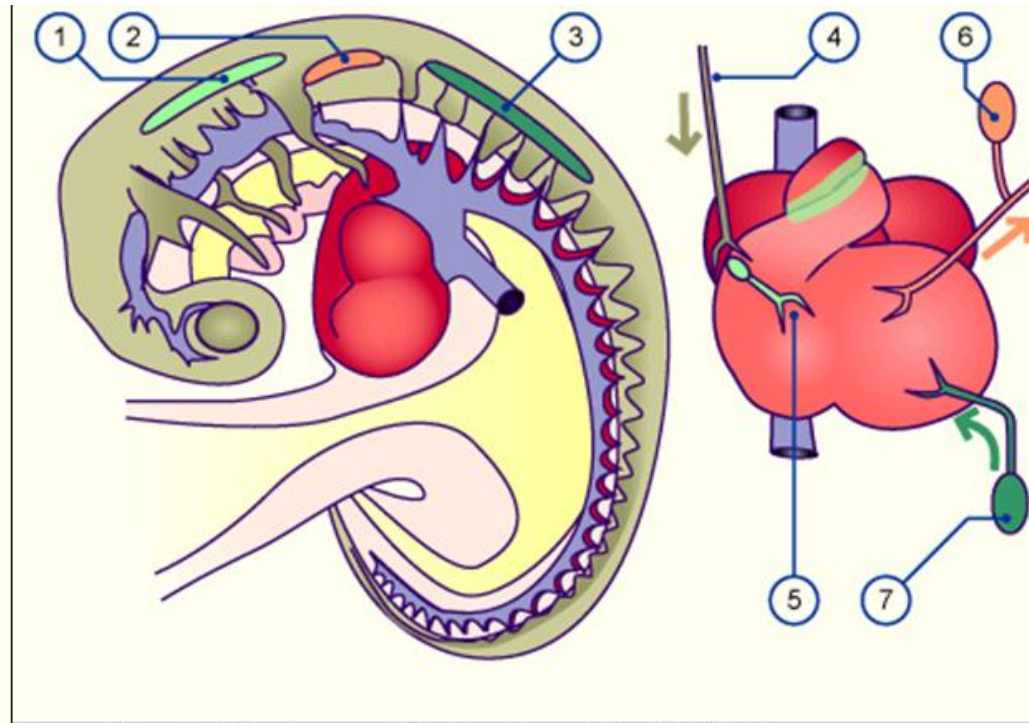


Cardiac neural crest cells



Cardiac neural crest cells

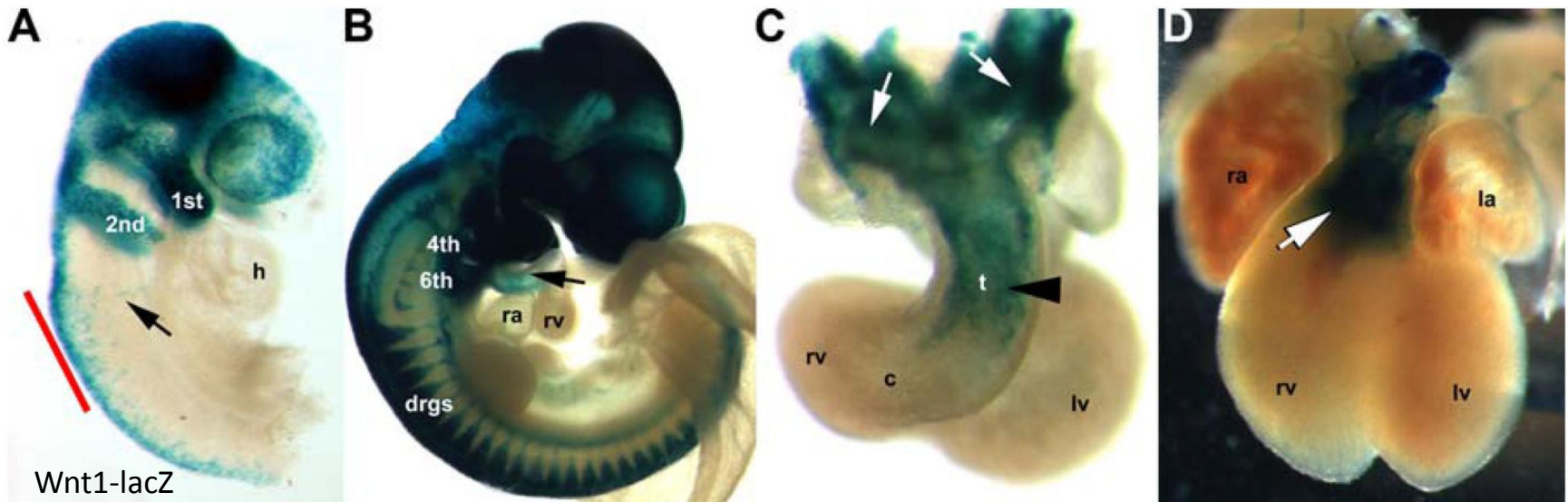
Cardiac innervation



- 1 Neural crest cells in the area of the rhombencephalon (hindbrain), parasympathetic, efferent
- 2 Ectodermal placode of the nodose ganglion (parasympathetic, afferent)
- 3 Neural crest cells of the trunk forming sympathetic ganglia (efferent)
- 4 First order parasympathetic neurons from cranial neural crest cells (1) contained in the vagus nerve (X)
- 5 Second order parasympathetic neurons from the cranial neural crest cells (1) that immigrate directly into the heart
- 6 Sensory ganglia that originate from the ectodermal placode of the nodose ganglion (2)
- 7 Sympathetic nerve fibers that originate from sympathetic ganglia of the trunk

Note also: cardiac outflow tract with neural crest cell material (1) migrated via the pharyngeal arches

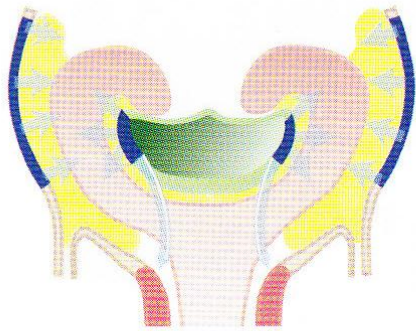
Cardiac neural crest cells



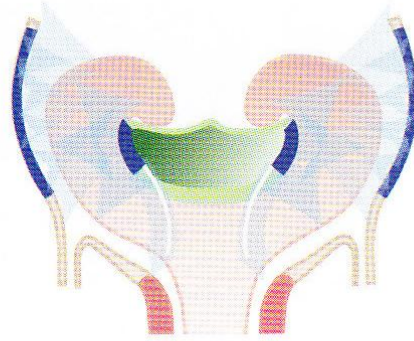
- CNC cells express Wnt1, Pax3, Cx43 during migration + colonization of the heart
- Wnt-signals: CNC induction, cell polarity
- Adhesion molecules (Cadherins) + Gap junction Cx43 provide „communication“ and interaction with their surrounding tissues during migration
- Pax3 is required for development of multiple neural crest lineages but is generally downregulated when NC cells leave the neural tube (maintains the undifferentiated state of mesenchymal NC cells)

Cardiac neural crest cells

CNC modulates endodermal FGF signals

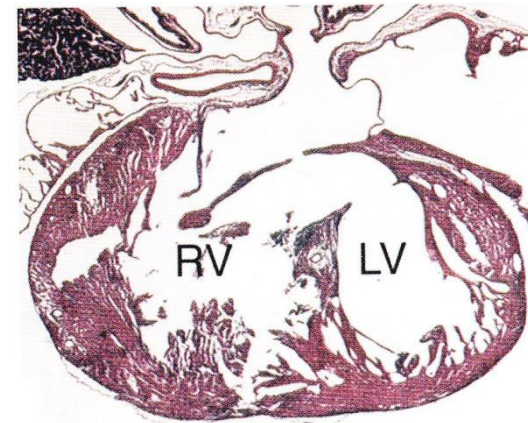


Normal



Cardiac neural
crest-ablated

Neural crest ablation



Ventricular septal defects

- Premigratory CNC ablation causes myocardial dysfunction
- suggestion: prolonged release of FGF signals by the pharyngeal endoderm
- these FGF signals are proposed to suppress myocardial development and alter myocardial proliferation/ differentiation
- Endocardium might have indirect negativ effects on myocardial maturation as a consequence of absent CNC colonization

summary

- Cardiac neural crest cells delaminate from the anterior neural tube between the otocyst and the 4th somite
- CNCs colonize the pharyngeal arches and the cardiac outflow tract where they differentiate into vascular smooth muscle covering the arterial endothelium
- Condensed CNCs form the aortic-pulmonary septation complex which modulates the septation of the arterial truncus into the aortic and pulmonary trunc
- CNCs contribute to the formation of the semilunar valves
- CNCs colonize the venous pole and provide cardiac innervation
- Pax3 is important for specifying neural crest lineages, blocks differentiation and is downregulated upon neural crest migration into the heart
- Adhesion and gap junction molecules provide interaction of CNCs with their surrounding tissue during migration
- CNCs modulate endodermal FGF8 signalling, thereby facilitating myocardial proliferation and differentiation

literature

- Epstein, J.A., Li, J., Lang, D., Chen, F., Brown, C.B., Jin, F., Lu, M.m., Thomas, M., Liu, E., Wessels, A., Lo, C.W., 2000. Migration of cardiac neural crest cells in Splotch embryos. *Development* 127, 1869-1878
- Kirby, ML., Waldo, K.L., 1995. Neural Crest and cardiovascular patterning. *Circ. Res.* 77,211-215
- Snider, P., Olaopa, M., Firulli, A.B., Conway, S.J., 2007. Cardiovascular development and the colonizing neural crest lineage. *TSW Dev + Emb.* 2, 88-111

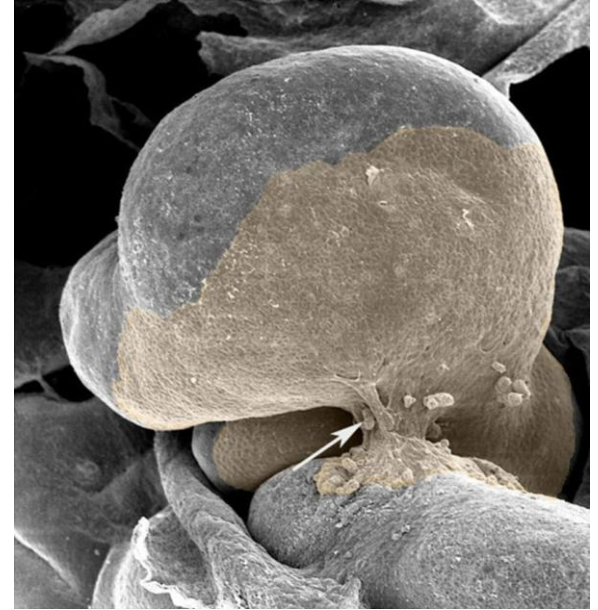
II Proepicardial Cells

1 Introduction

2 Asymmetrical PE development

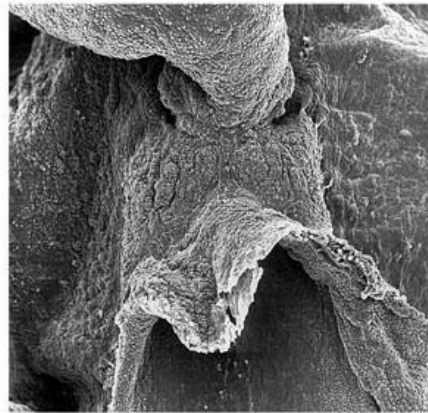
3 The role of BMP and FGF

4 Neovascularization and regeneration

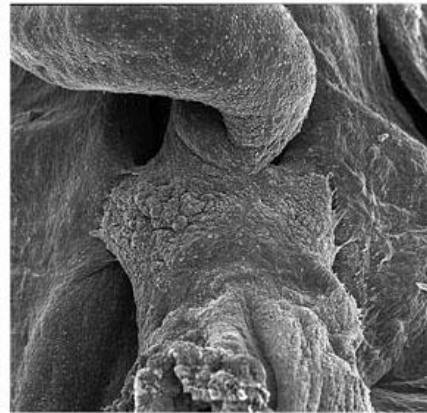


1. Introduction

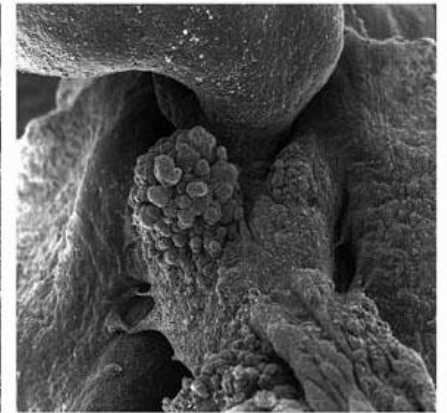
- The proepicardium (PE) develops on the sinus venosus of the heart



HH 14

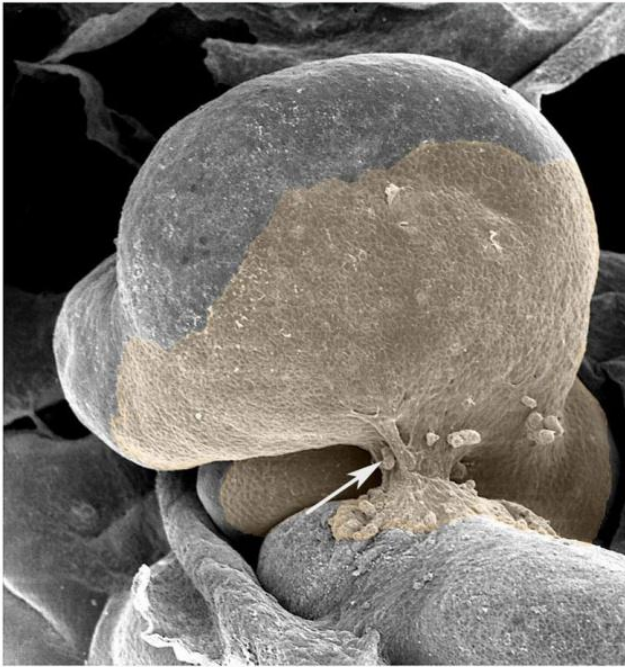


HH 15

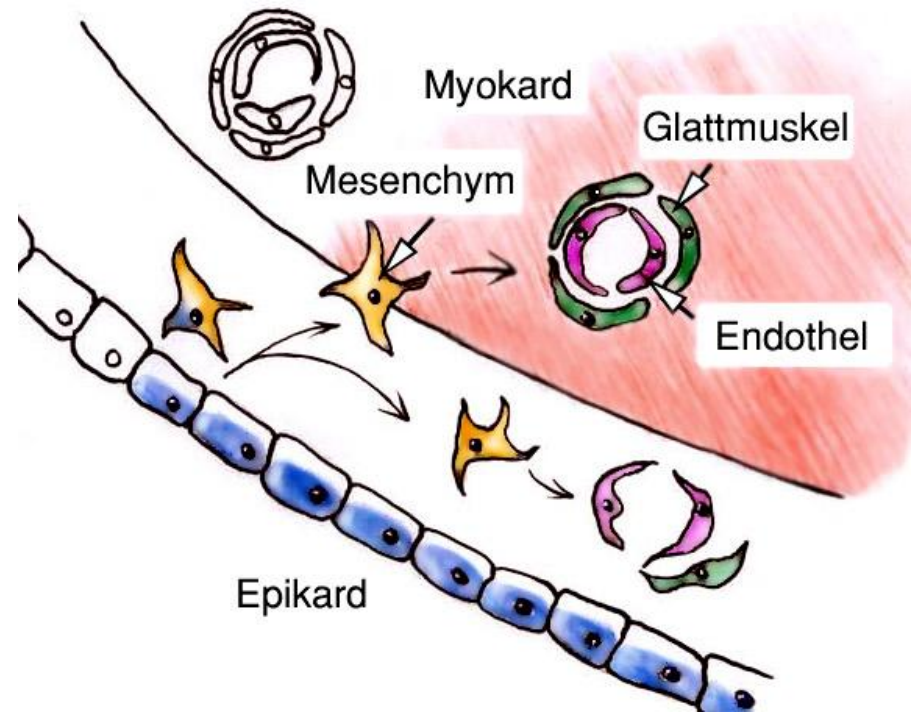


HH 16

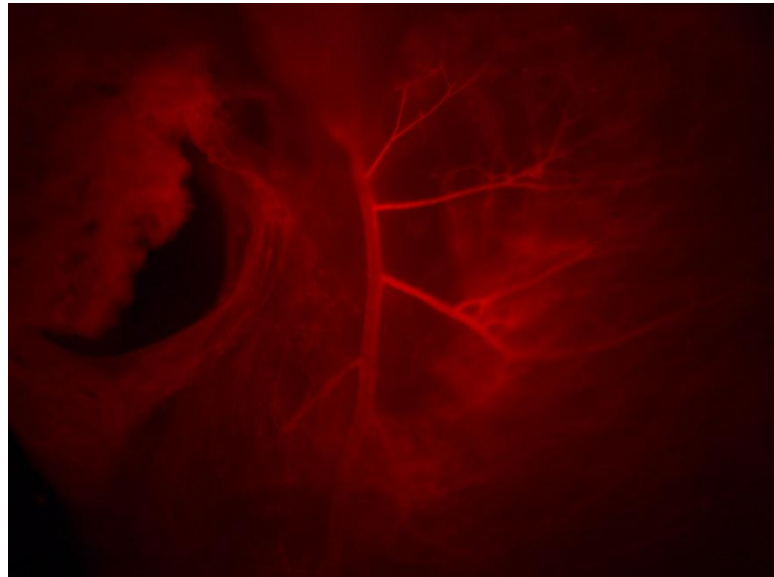
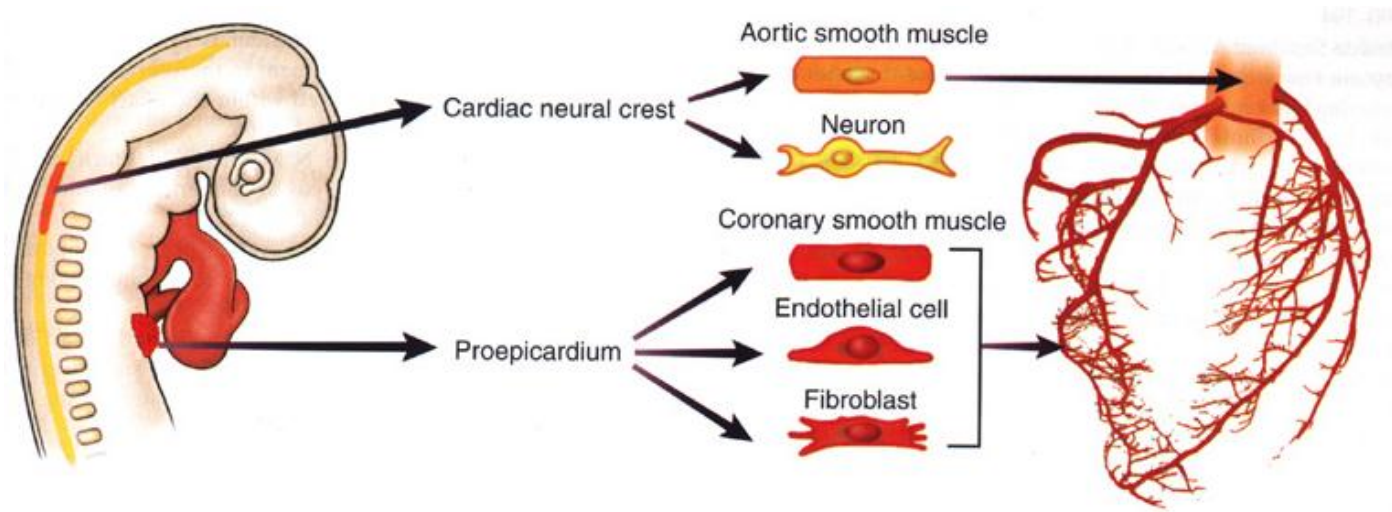
Epicardial cells invade the heart



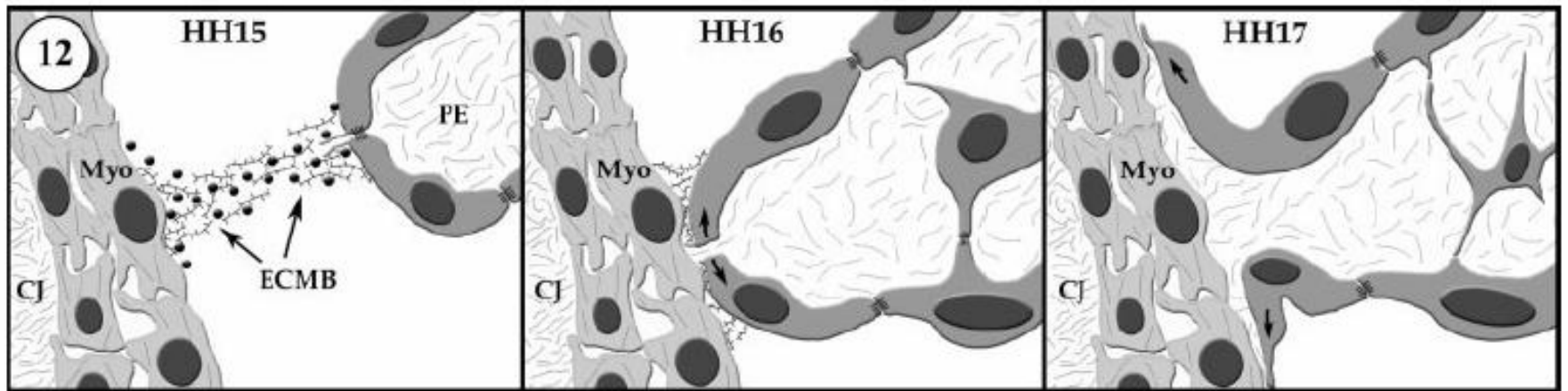
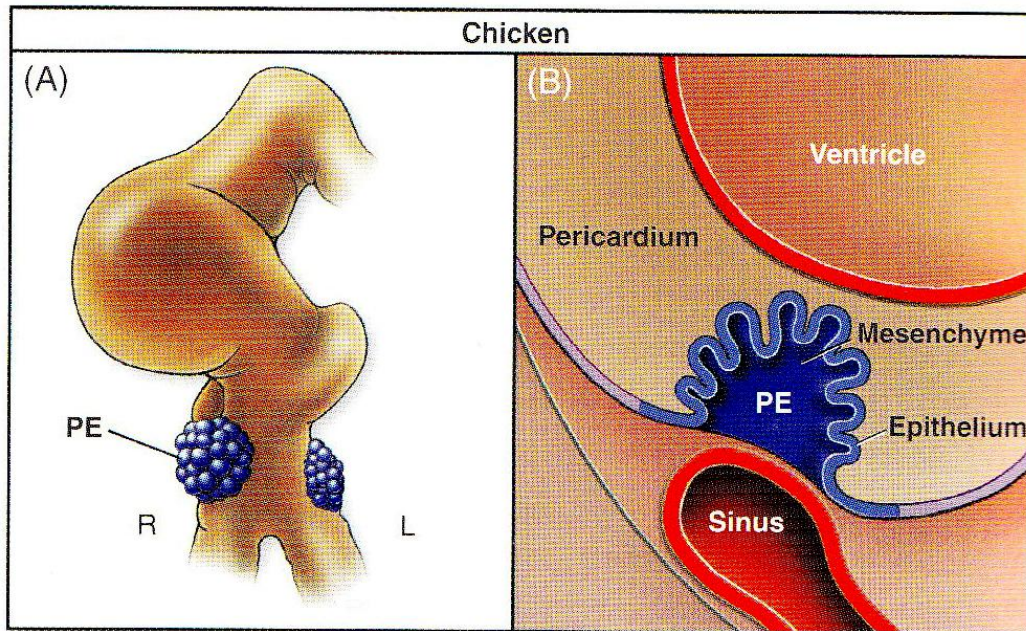
(Männer et al., 2000)



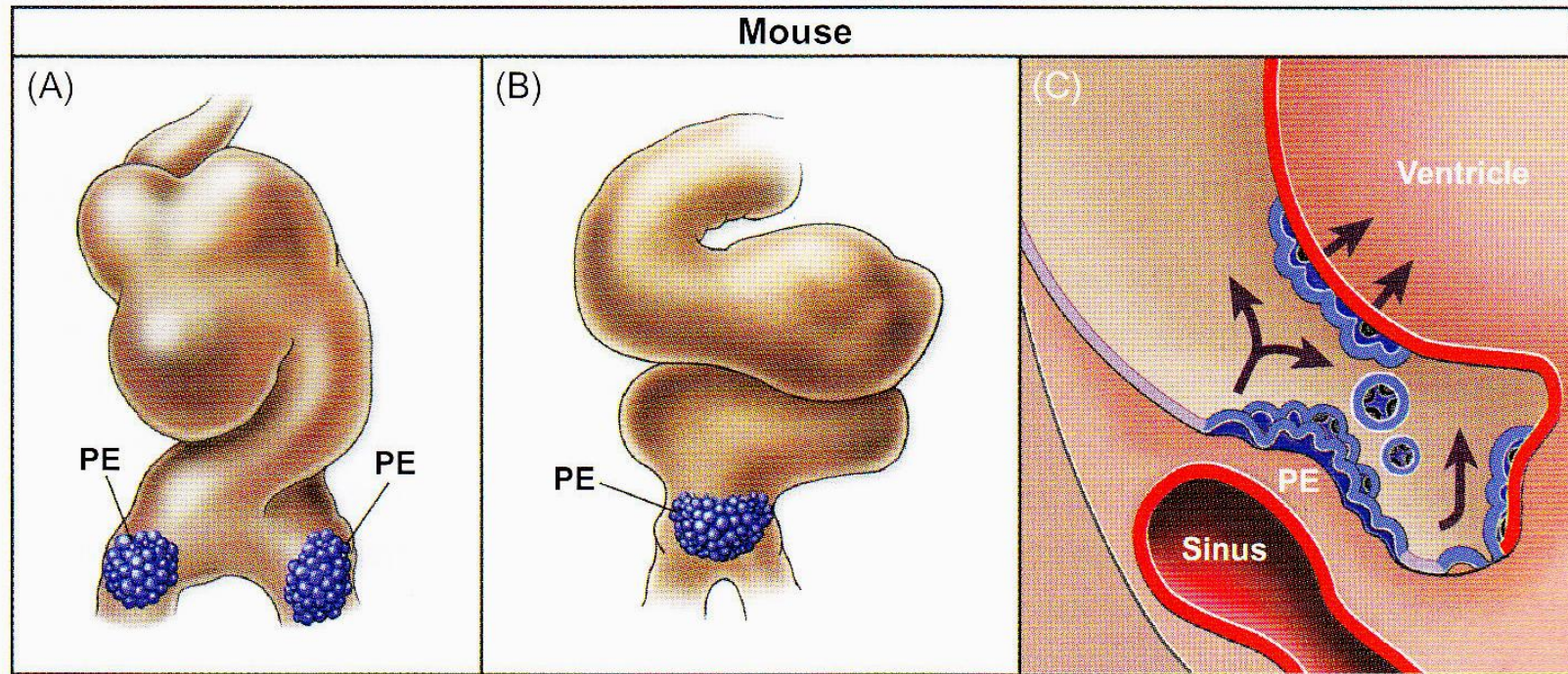
EPDCs (epicardial derived cells) differentiate into coronary blood vessels



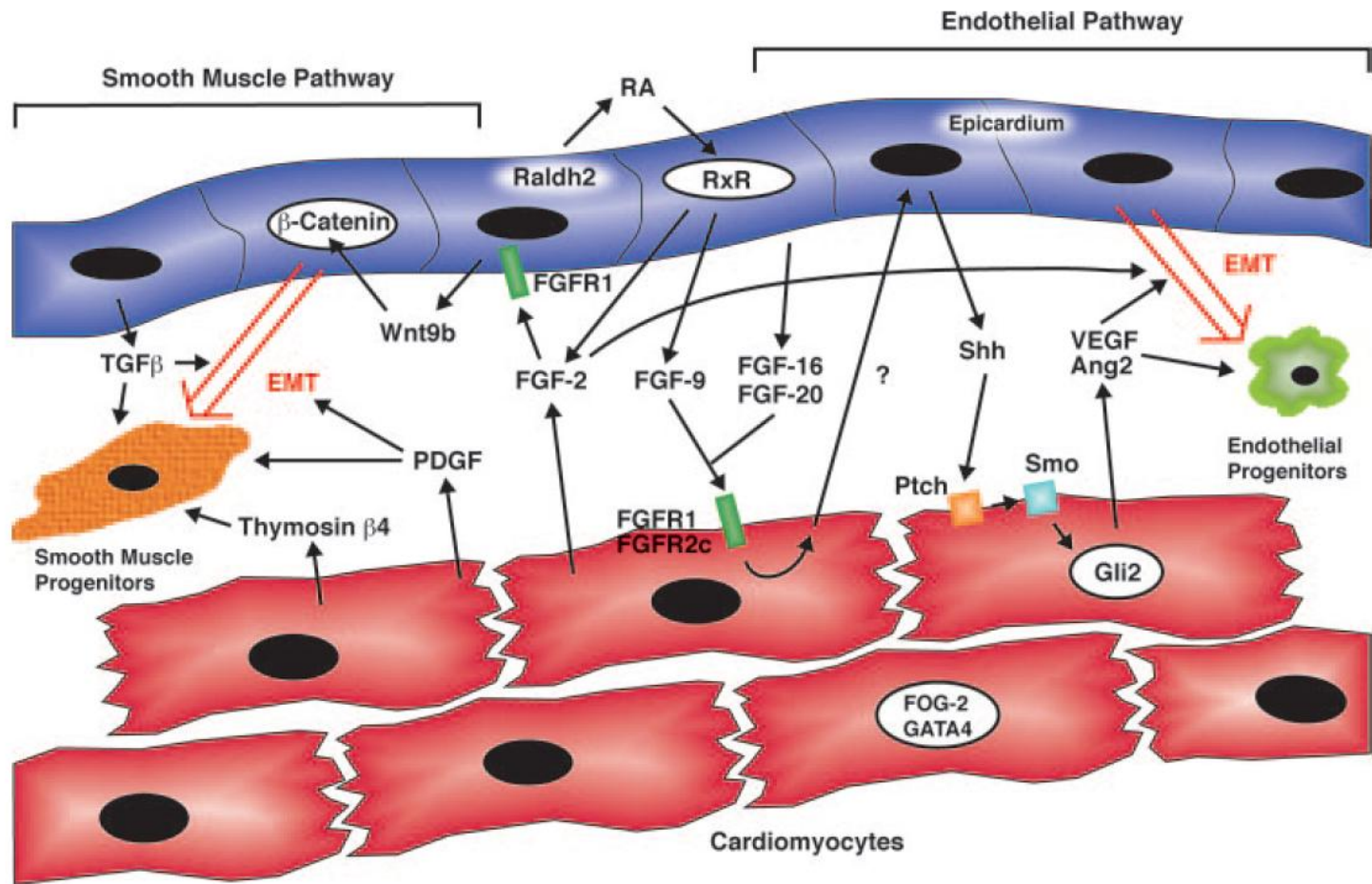
PE cells colonize the heart via a tissue bridge in the chicken embryo



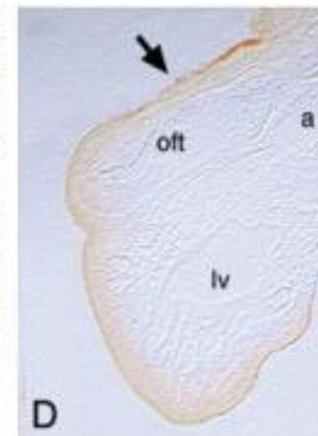
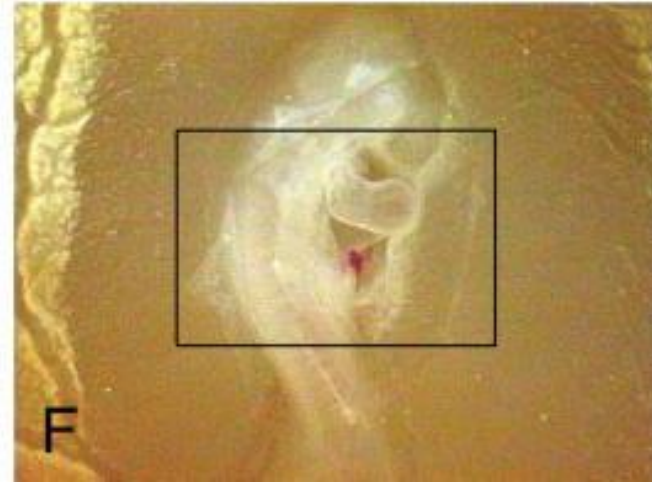
PE cells colonize the heart via proepicardial cysts in the mouse embryo



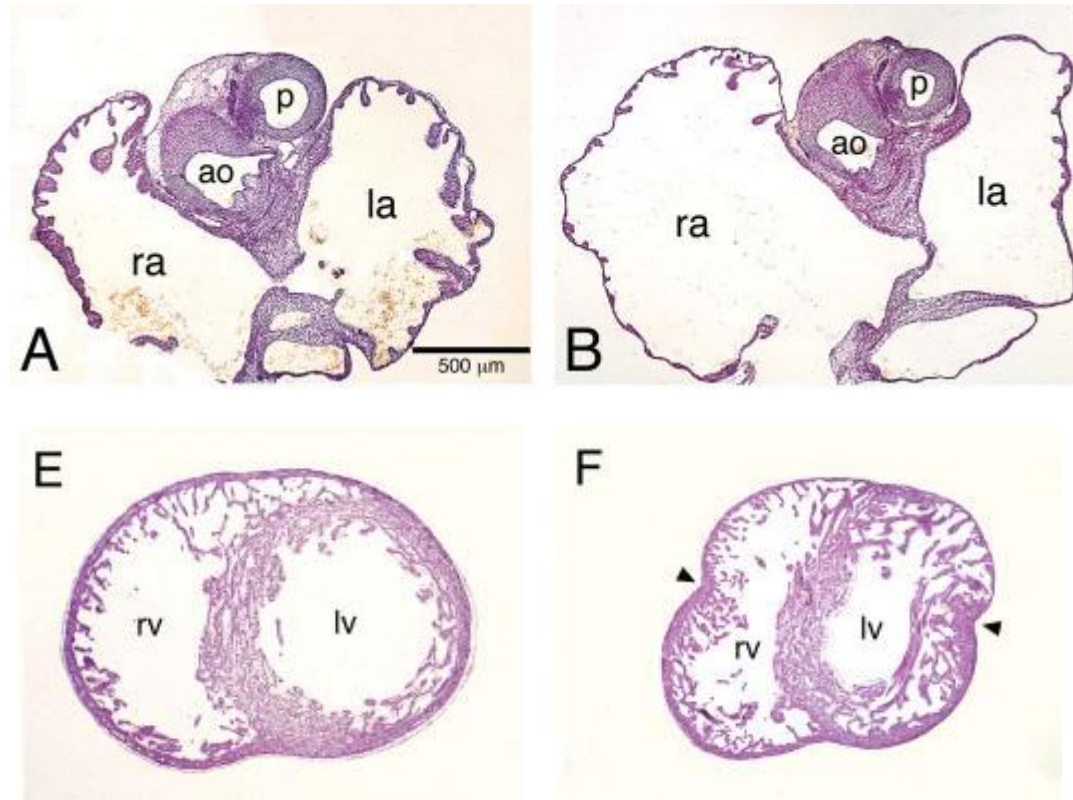
Epicardium-myocardium interactions



Loss of proepicardial cells leads to loss of the epicardium



Retarded myocardial growth without epicardium



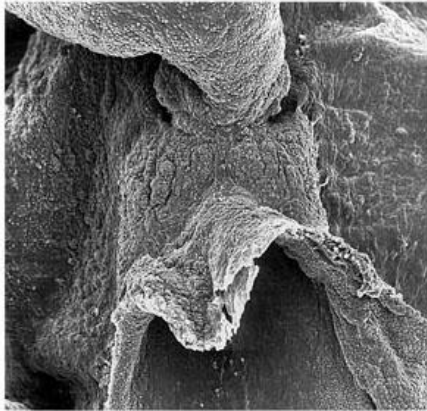
(Maenner et al., 2005)

Summary 1

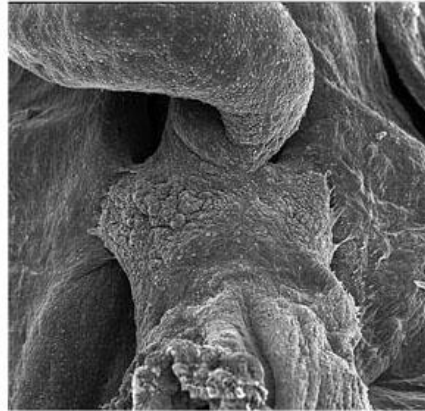
- The PE (proepicardium) develops on the venous pole of the heart and subsequently colonizes the heart to form the epicardium
- the process of colonization differs in chick and mouse embryos
- Epicardial cells invade the heart and differentiate into the coronary vasculature
- The myocardium is strongly dependent on interaction with the epicardium which promotes myocardial growth

2. Asymmetrical PE development

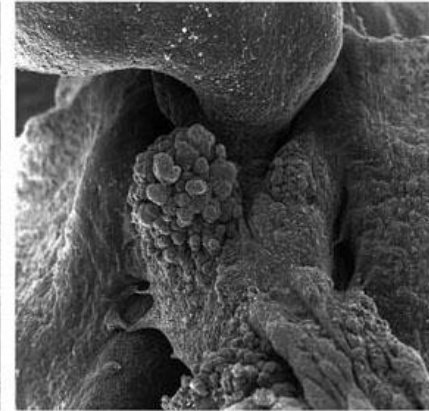
chick



HH 14

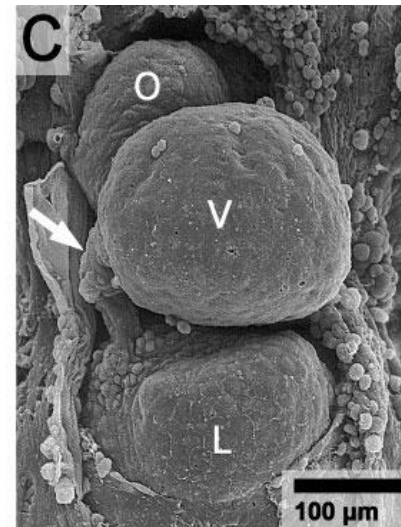
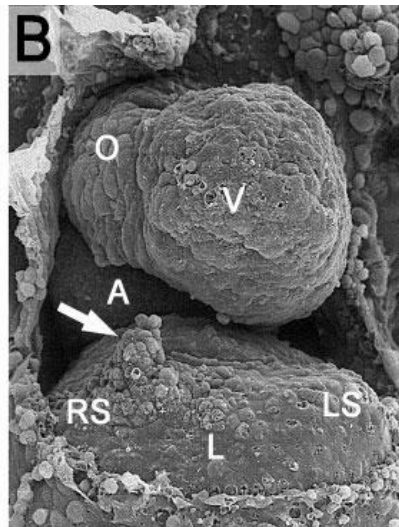
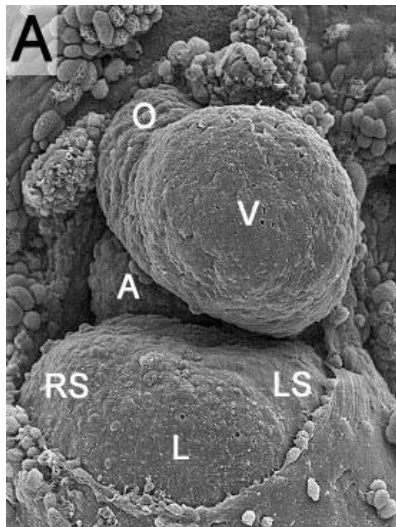


HH 15



HH 16

frog



The asymmetrical PE anlage

Vertebrata:

Agnatha

lamprey



Gnathostomata

Osteichthyes

zebrafish



Tetrapoda

Amphibia

xenopus



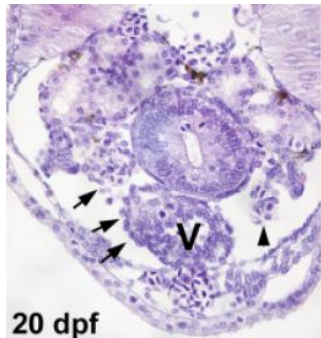
Aves

chick



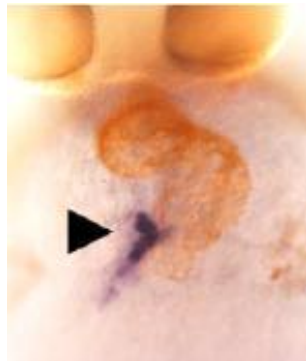
Mammalia

mouse

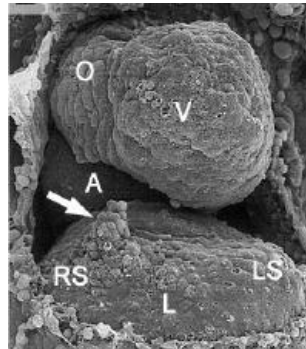


20 dpf

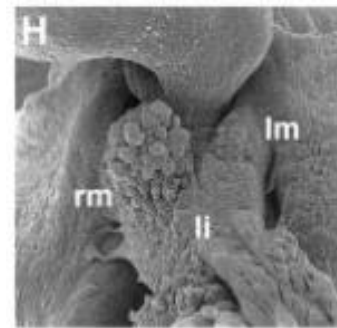
(Pombal et al., 2008)



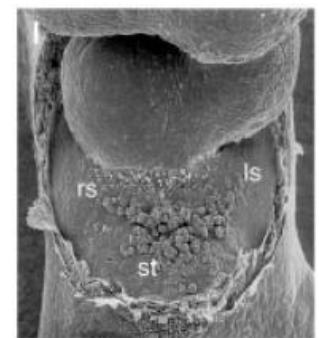
(Serluca, 2007)



(Jahr et al., 2008)

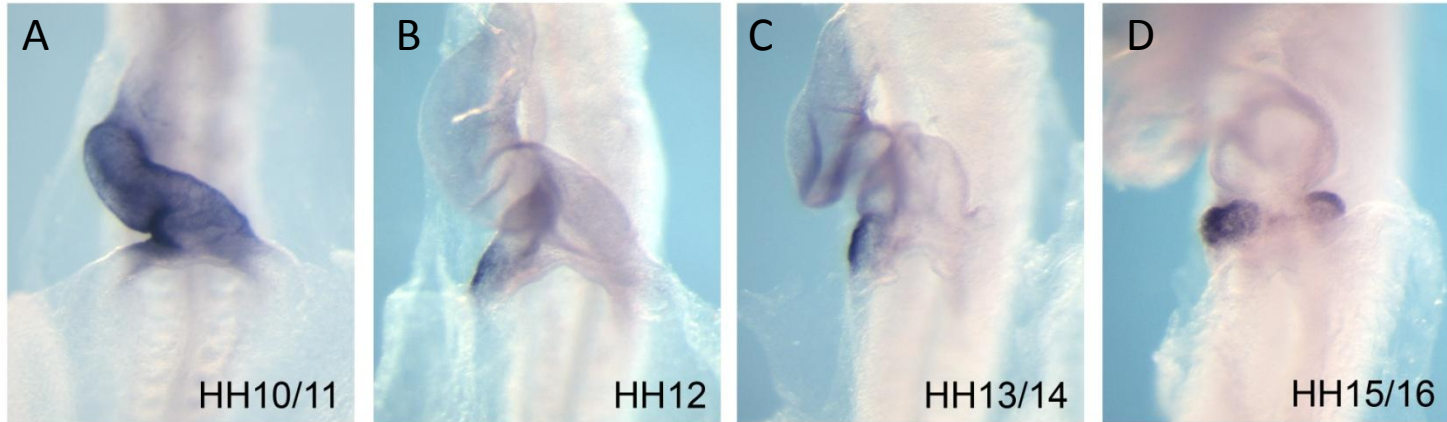


(Schulte et al., 2007)

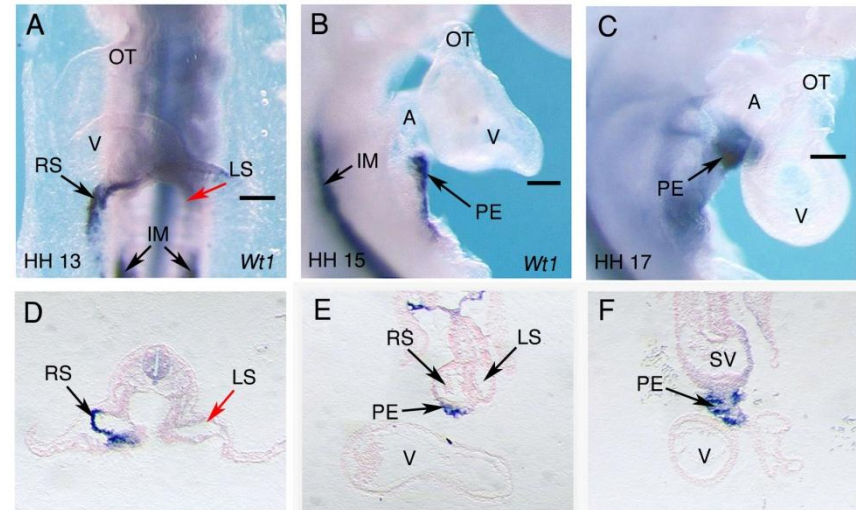
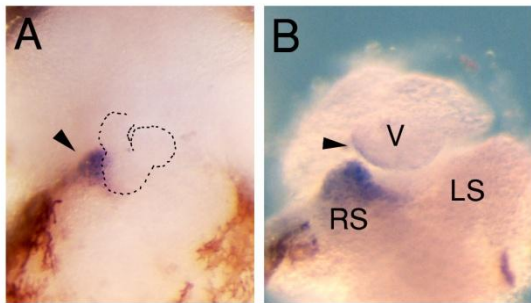


The expression of proepicardial marker genes

TBX18
chick



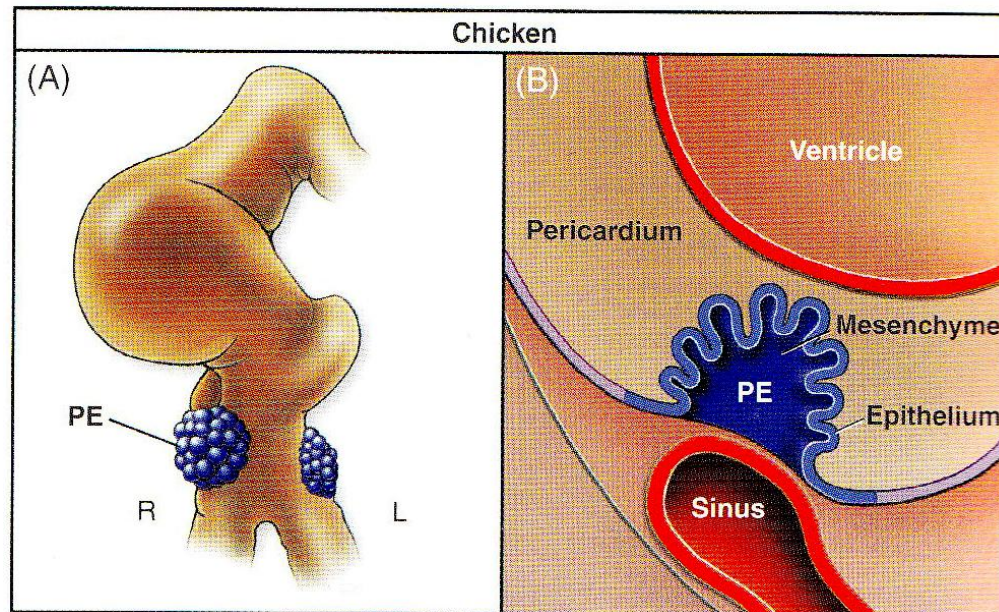
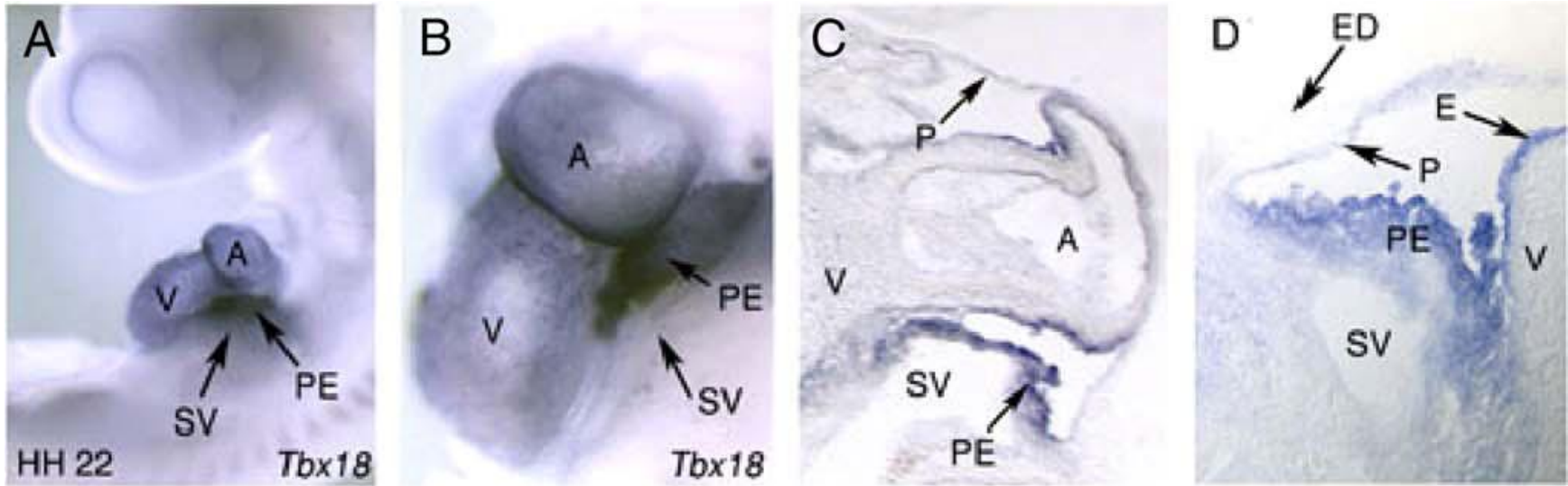
TBX18
Xenopus
(42h)

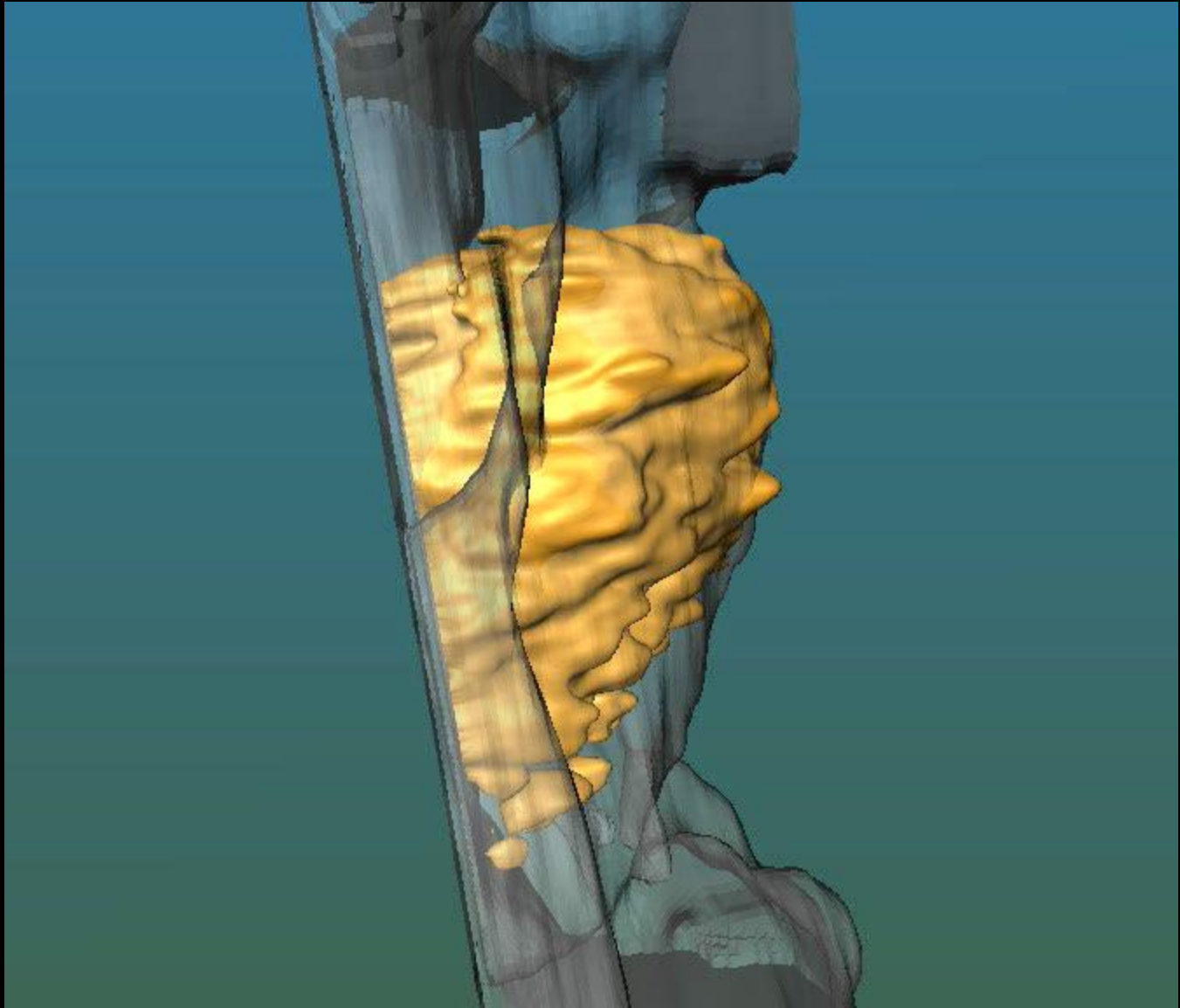


WT1 chick

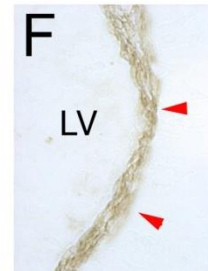
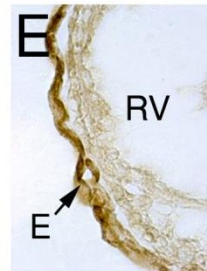
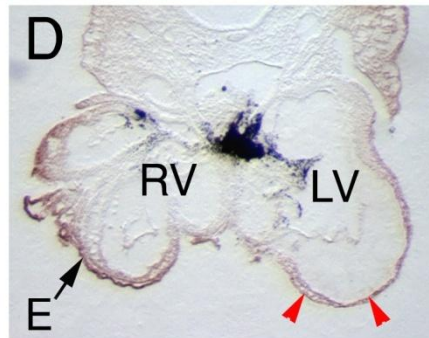
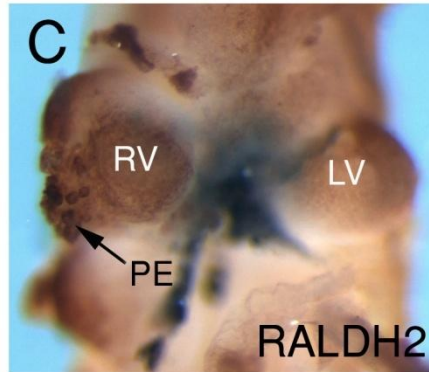
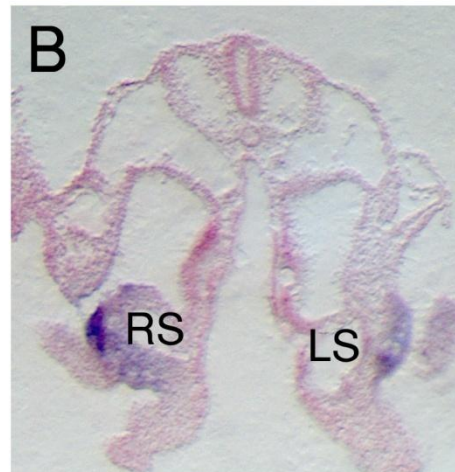
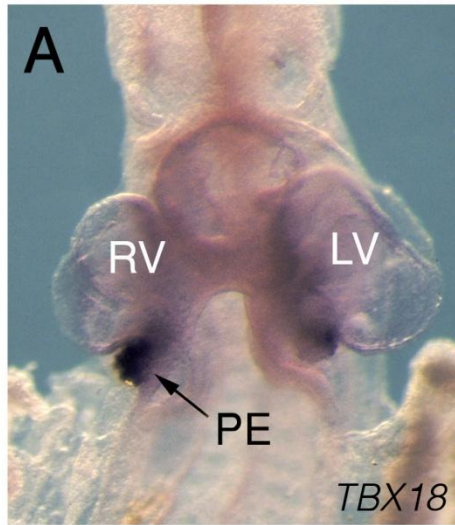
(Schlueter et al., 2006)

The expression of proepicardial marker genes





Cardia bifida embryos display only a right epicardium



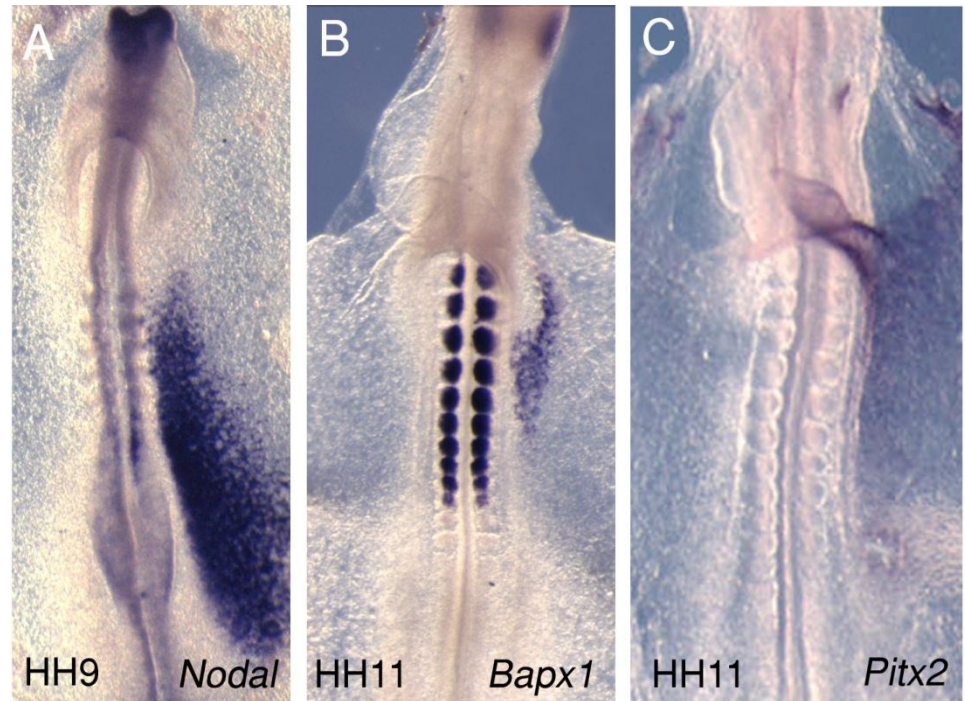
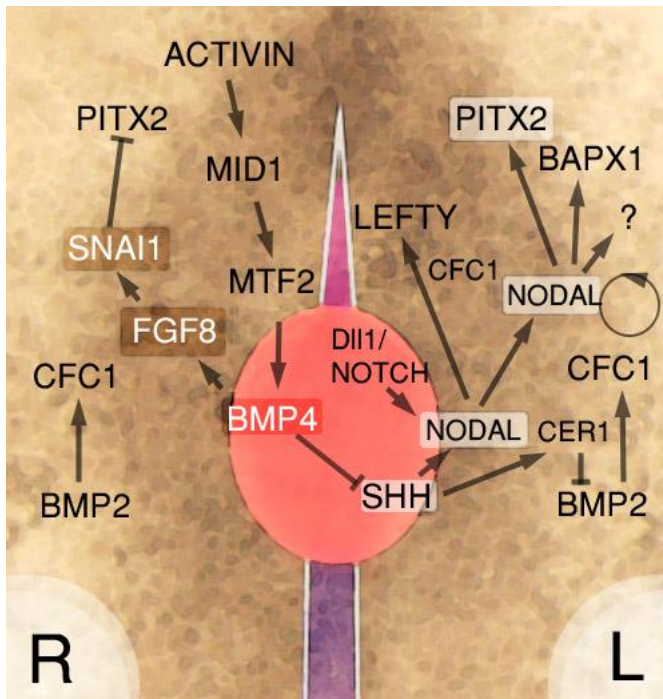
➤ The PE has a right identity

Question:

Is a left-sided repressive signal responsible for asymmetrical PE development?

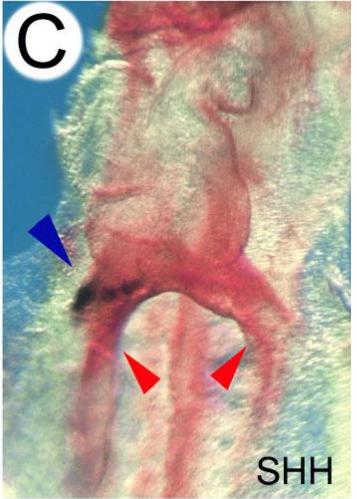
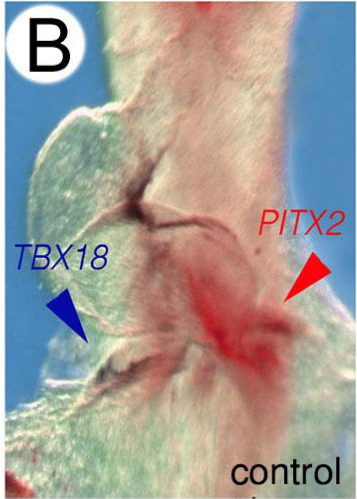
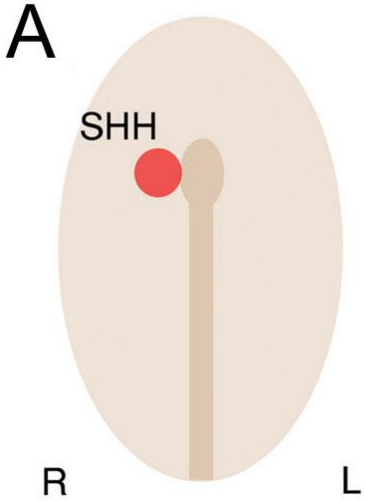
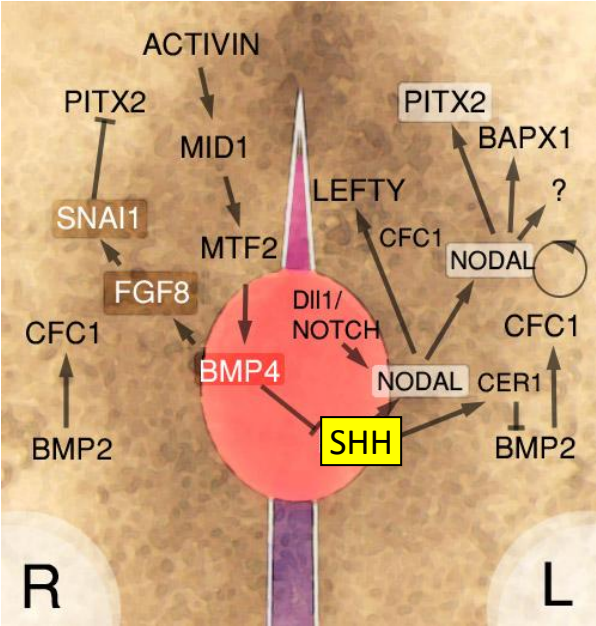
The Nodal/PITX2 pathway

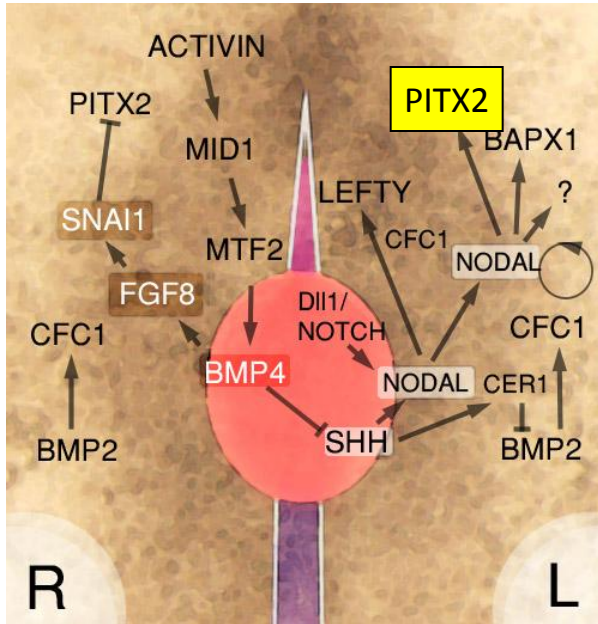
The conserved NODAL-PITX2 signaling pathway determines the left side



(Schlueter and Brand, 2007)

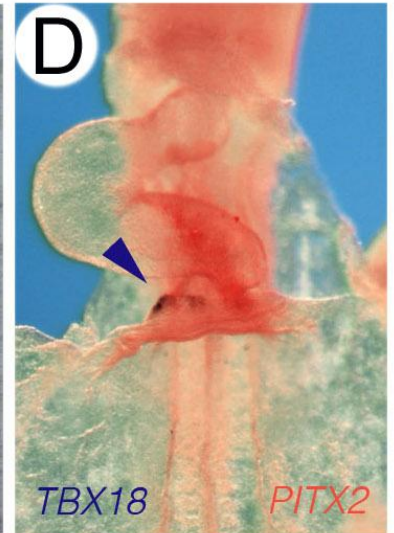
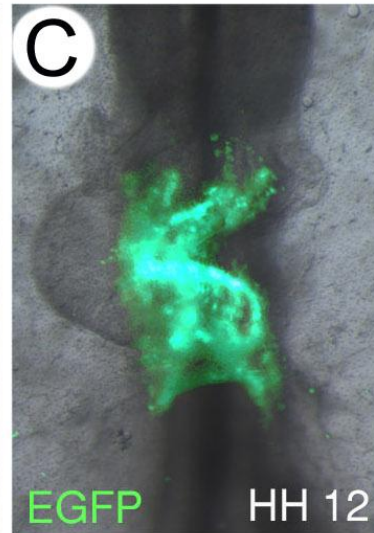
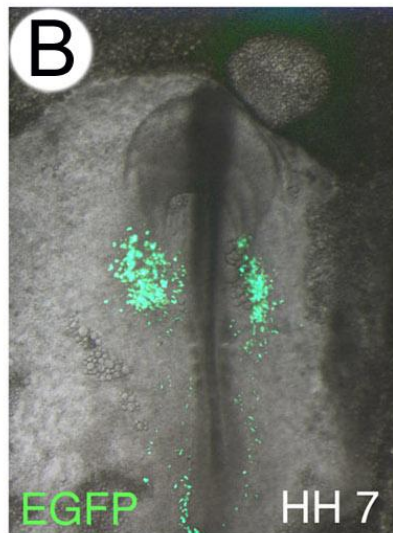
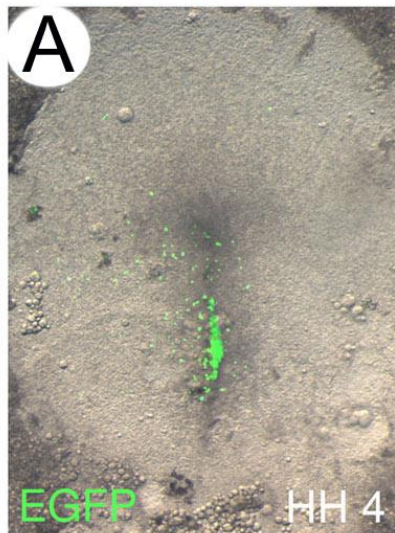
Gain-of-function of PITX2



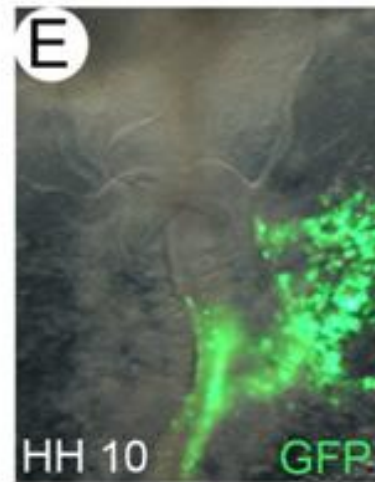
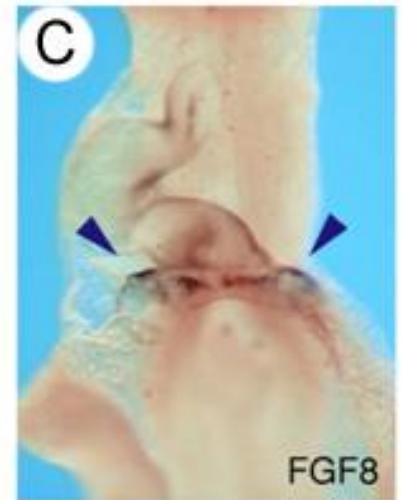
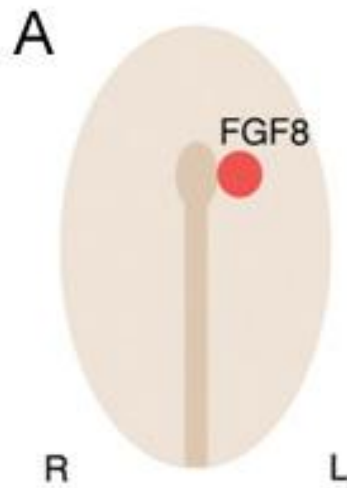
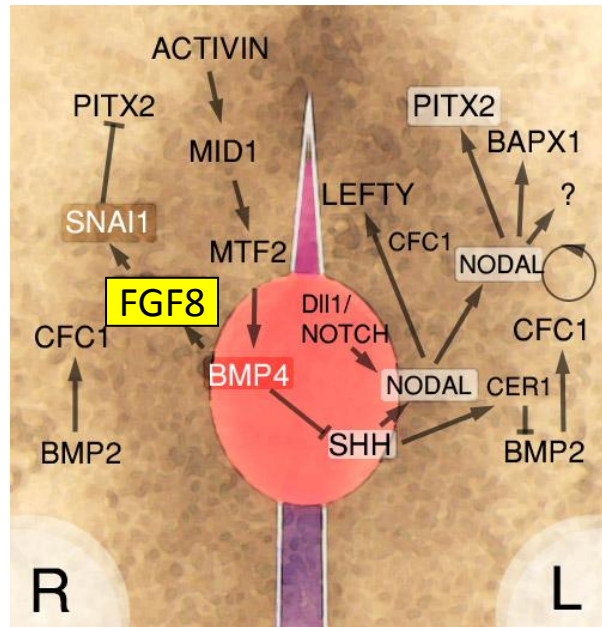


The overexpression of PITX2

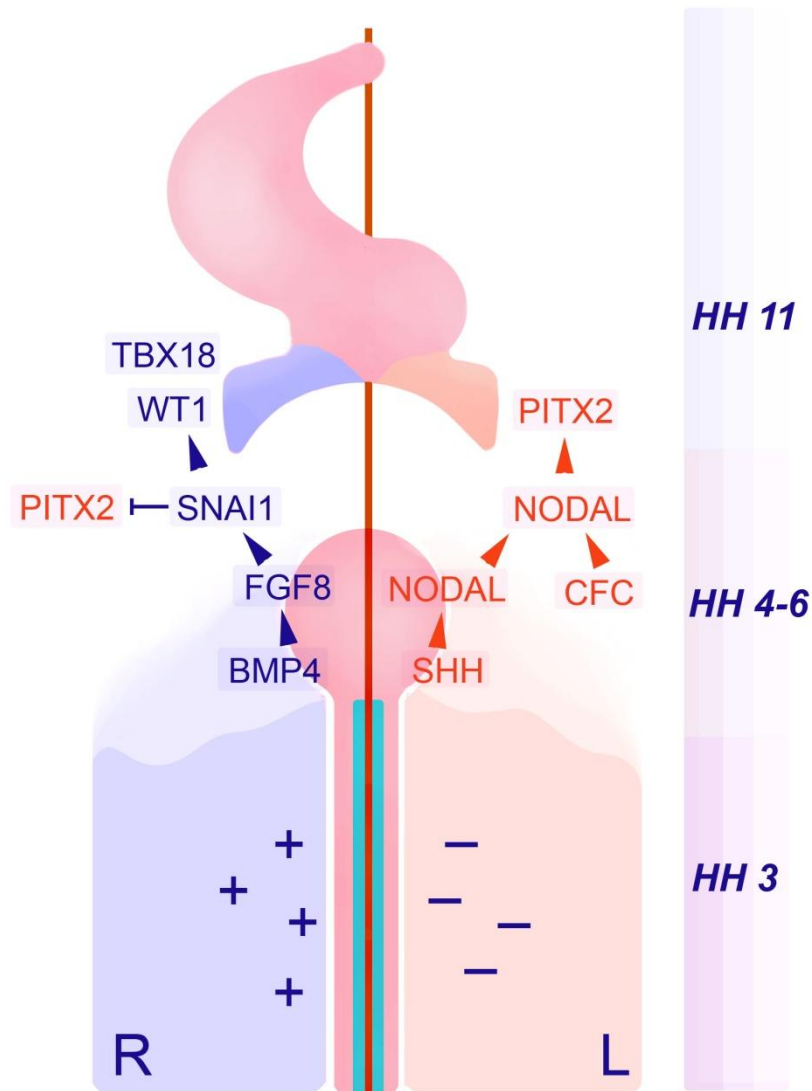
- Electroporation of sinuatrial progenitors with PITX2/RCAS construct
- No effect on TBX18
- the PE is lateralized independently of the NODAL-PITX2 signaling pathway!



Gain-of-function of FGF8/ SNAI1 leads to bilateral PE anlagen

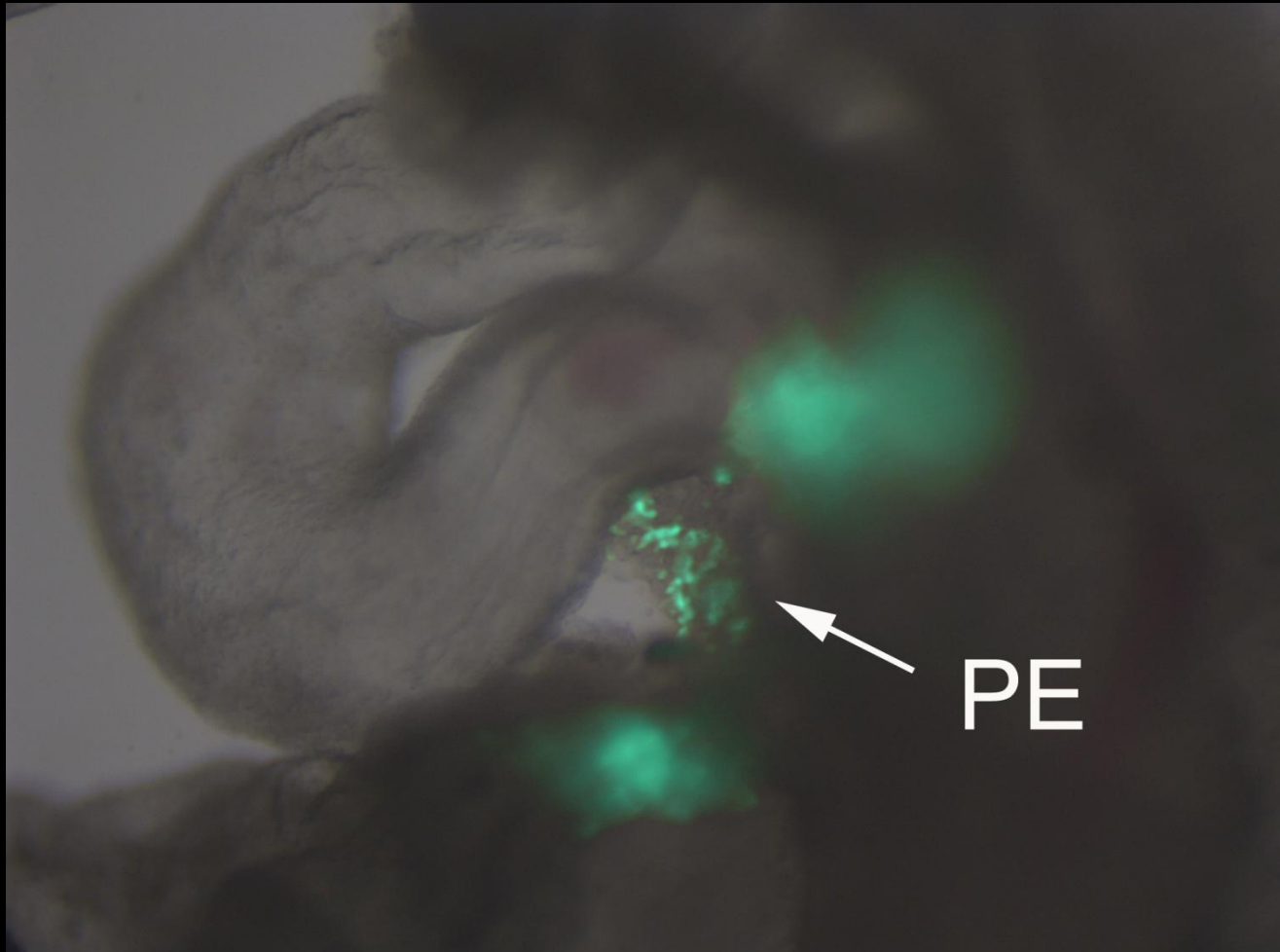


A rightsided FGF8/SNAI1 pathway induces proepicardial gene expression

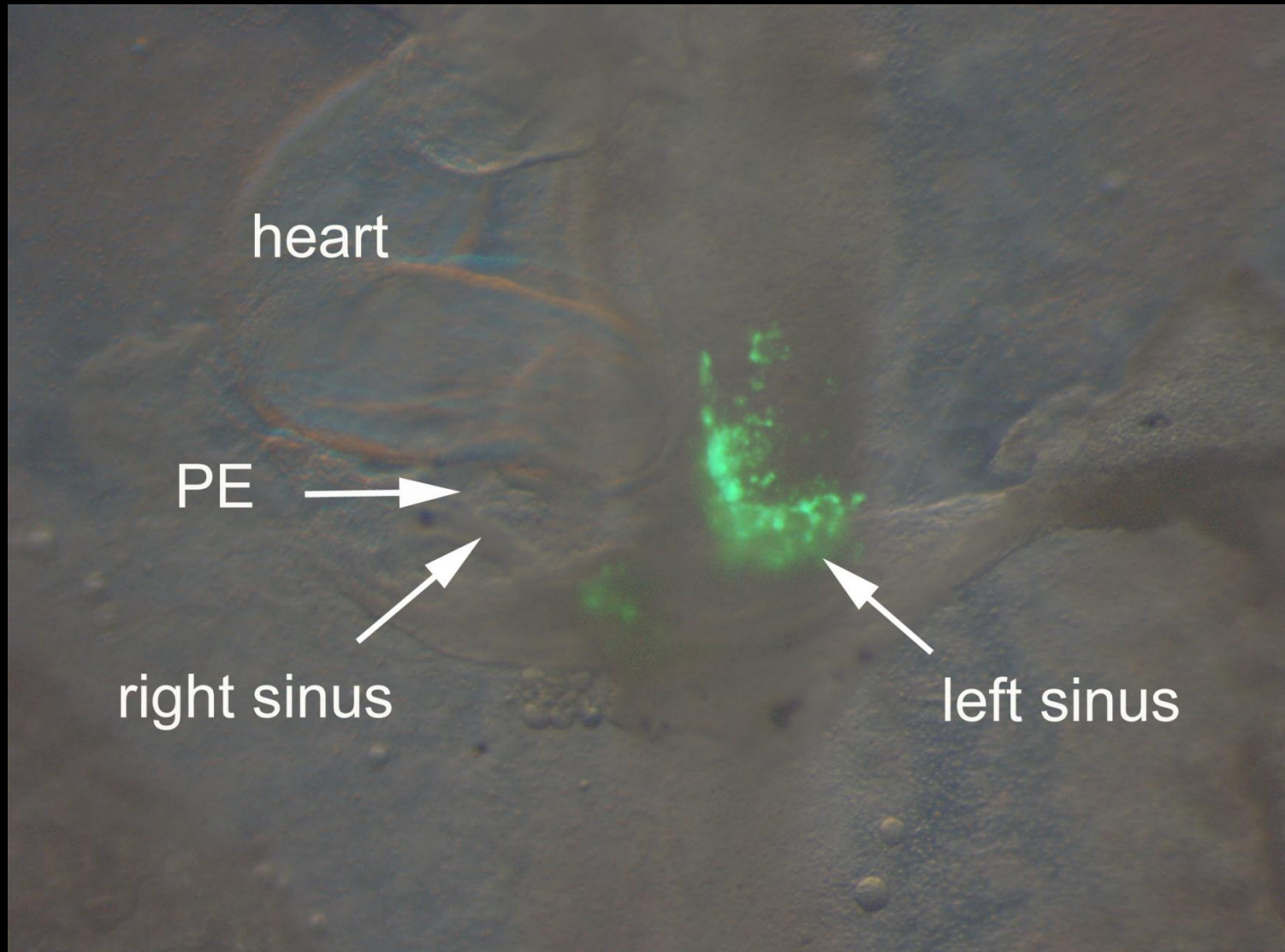


(Schlueter and Brand, 2009)

Transfection of proepicardial cells – gene delivery (live imaging)



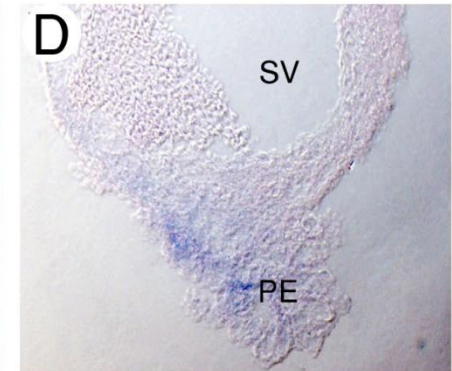
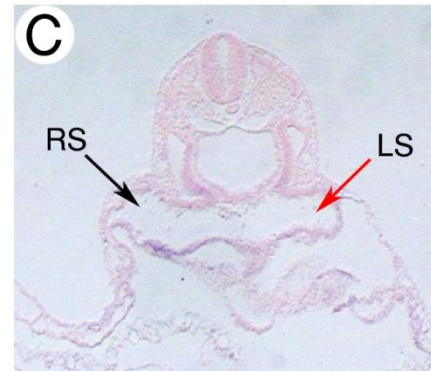
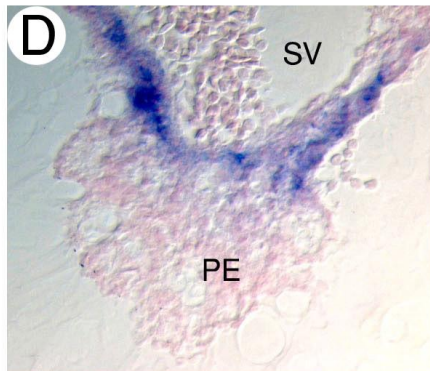
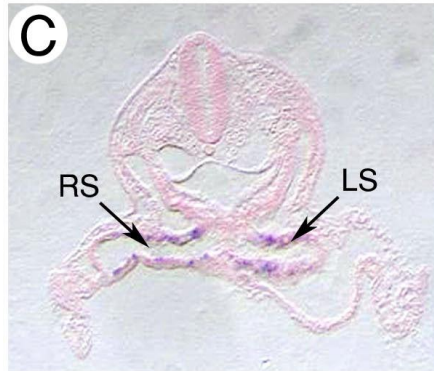
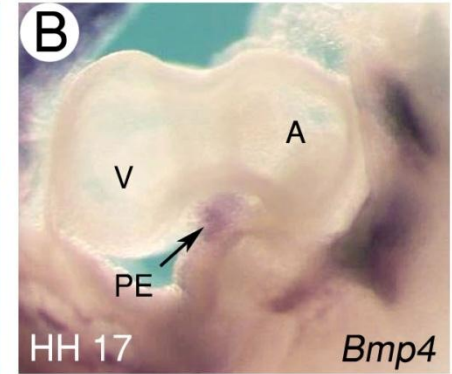
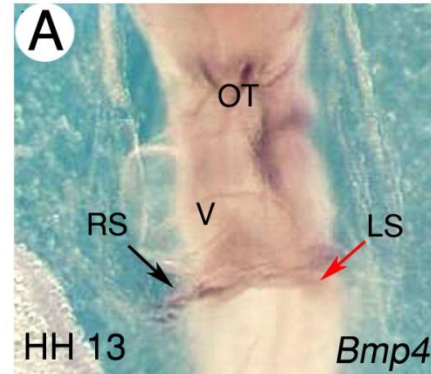
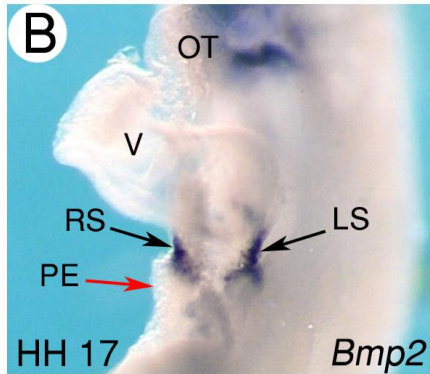
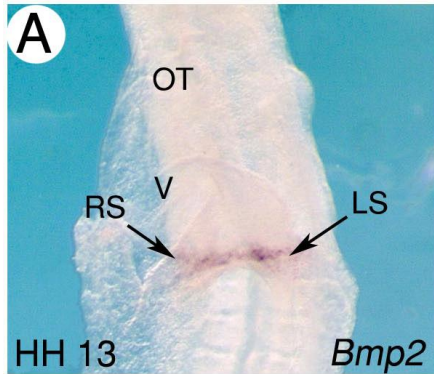
Induction of proepicardial gene expression programme on the left side



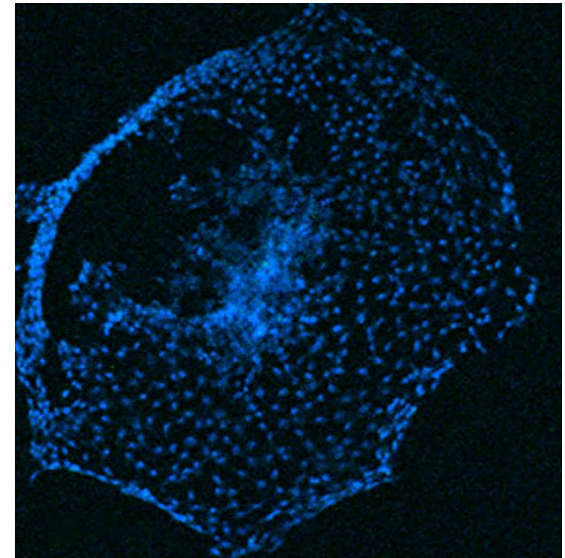
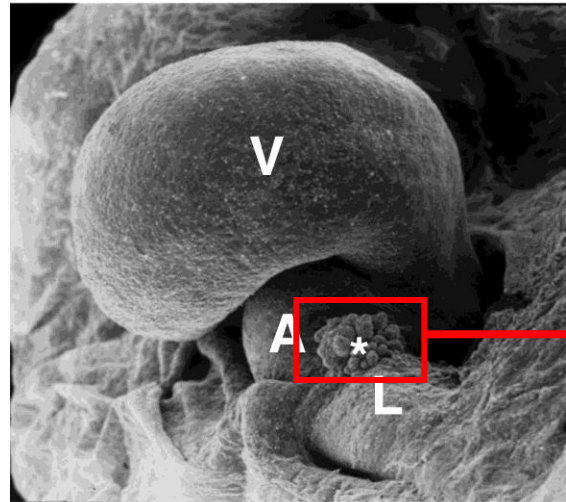
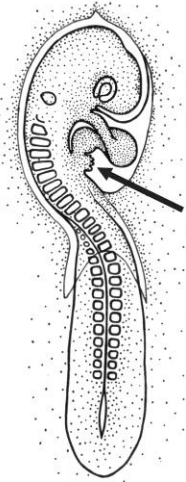
Summary 2

- The PE develops stronger on the right sinus venosus in lower vertebrates
- TBX18 and WT1 are expressed in the PE and the pericardium
- PE marker genes are not regulated by the NODAL/PITX2 pathway
- Instead, both marker genes are regulated by a rightsided FGF8/ SNAI1 pathway

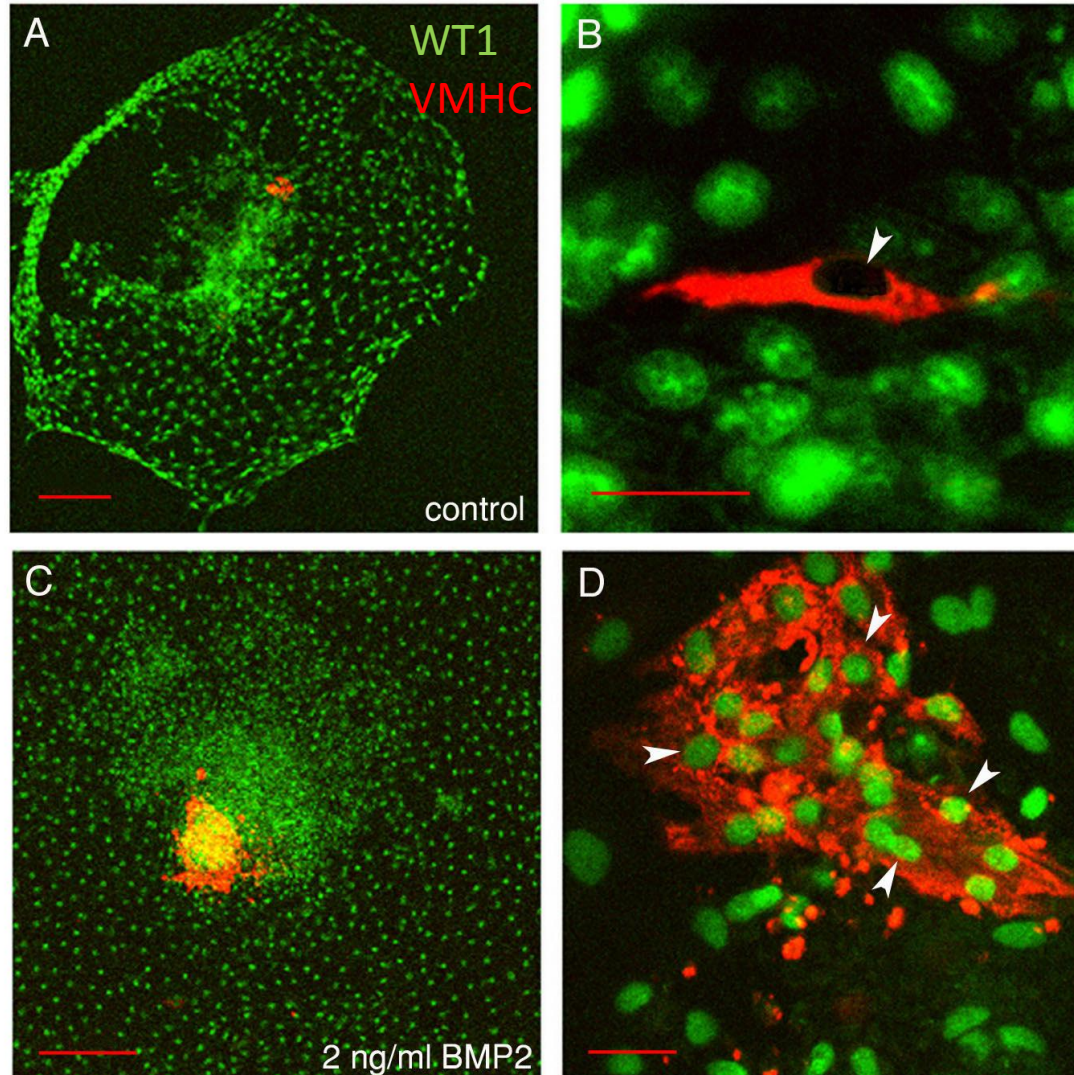
3. The role of BMP and FGF during PE development



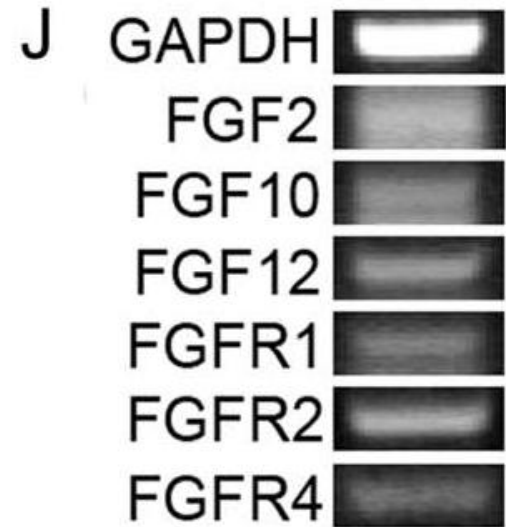
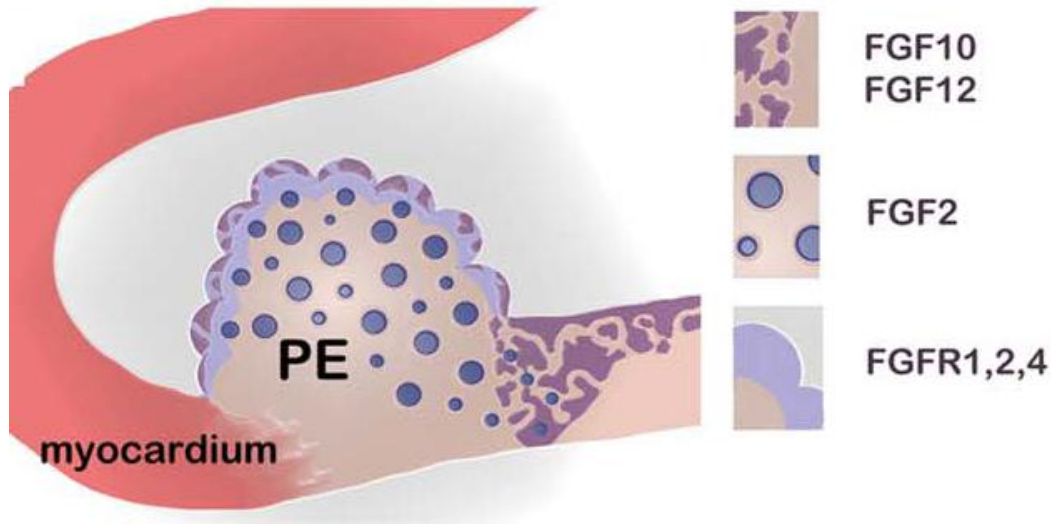
In vitro explant assay



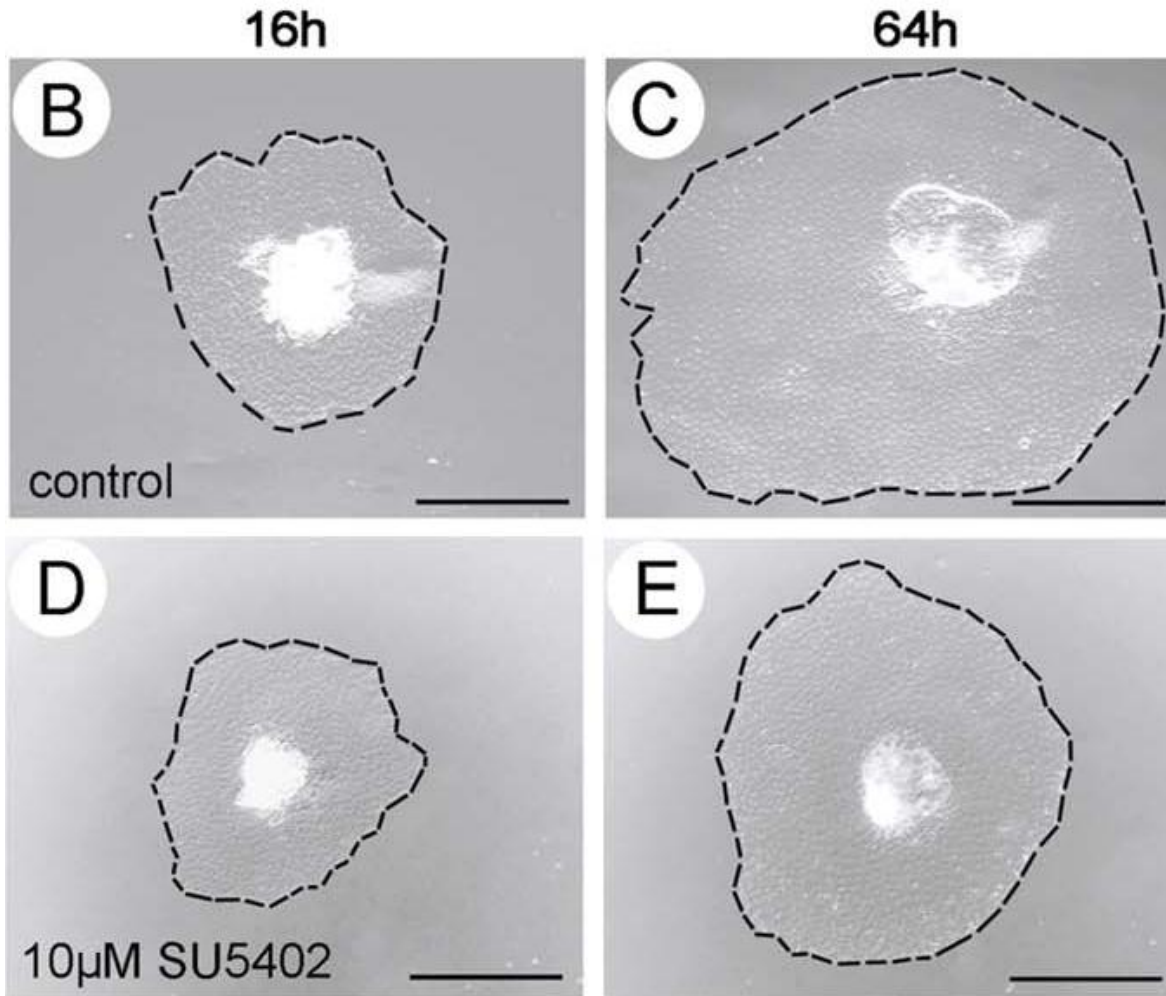
BMP promotes myocardial differentiation



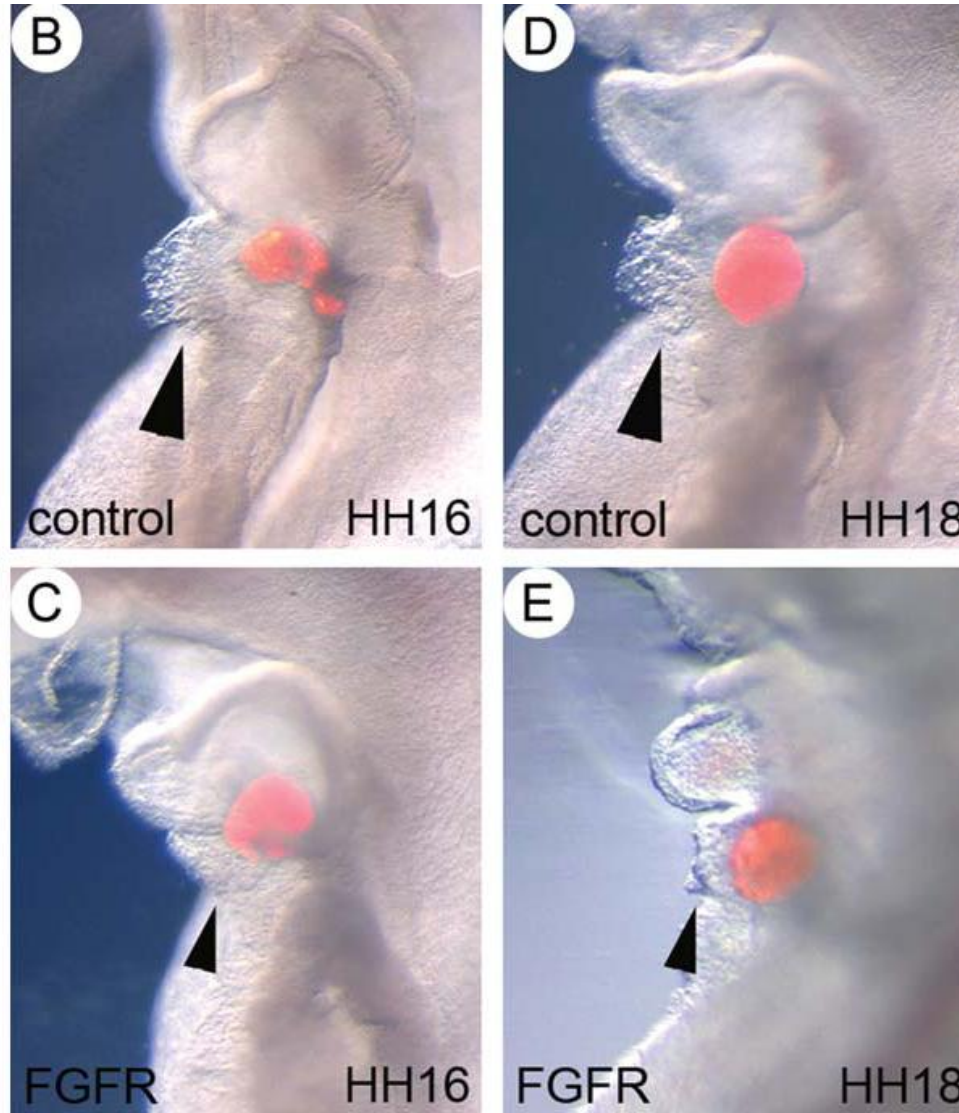
Proepicardial expression of FGF ligands and receptors



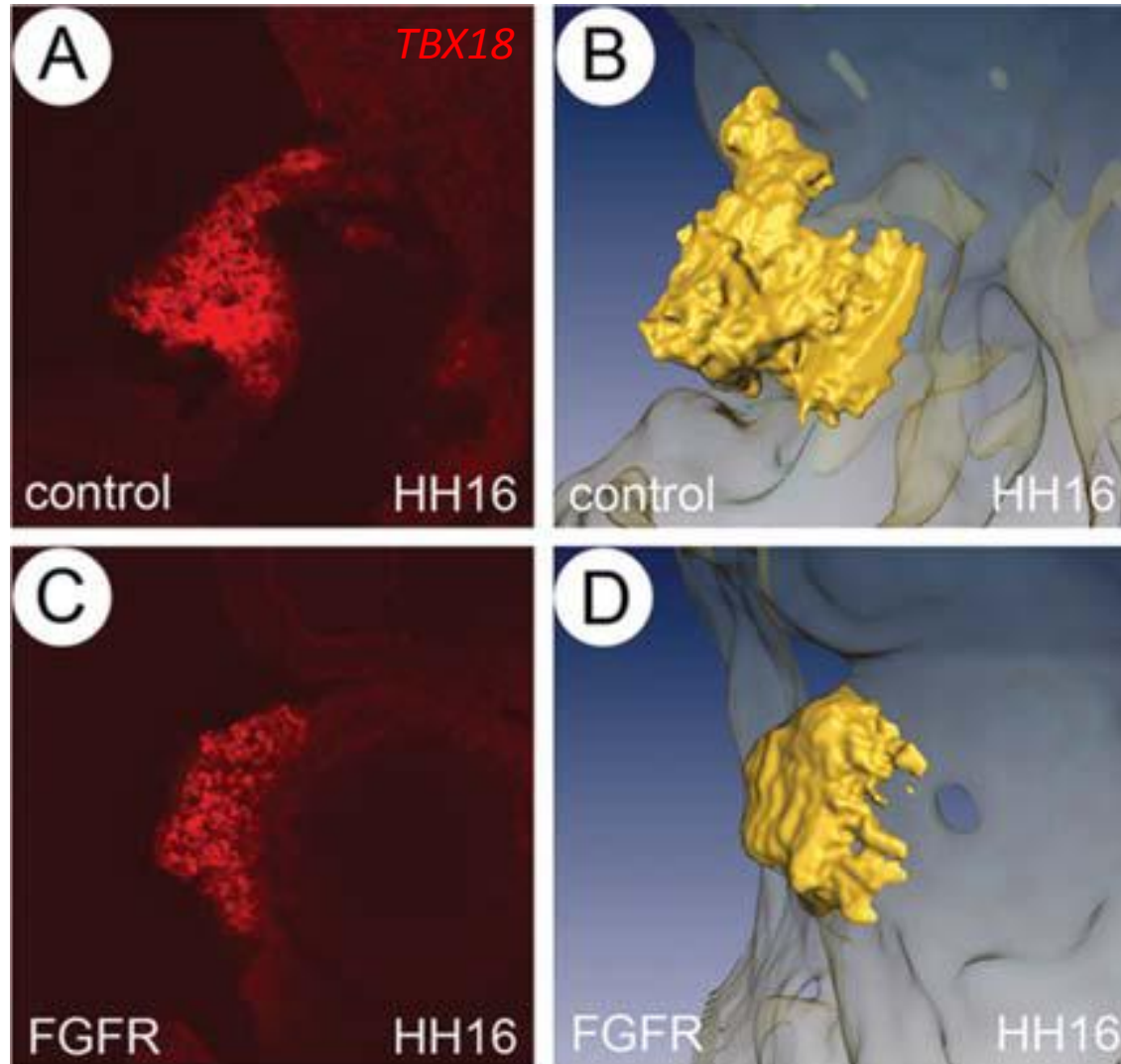
Loss of FGF signalling leads to retarded growth and cell death in PE explants



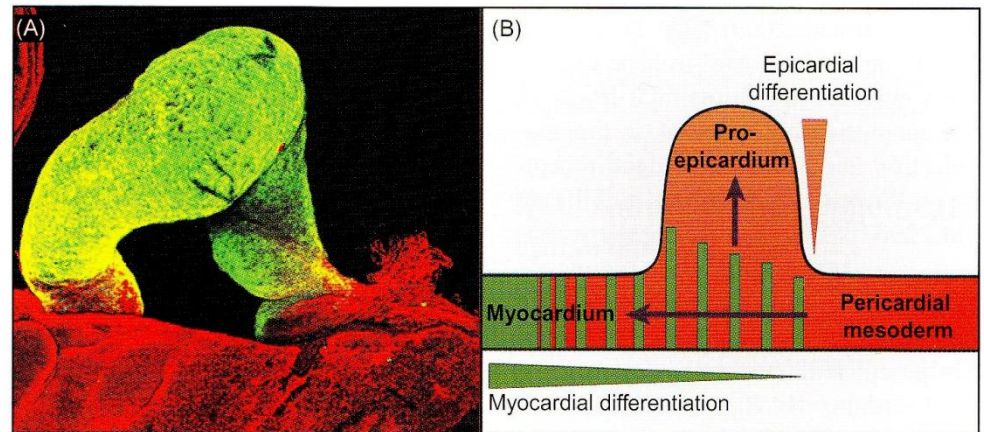
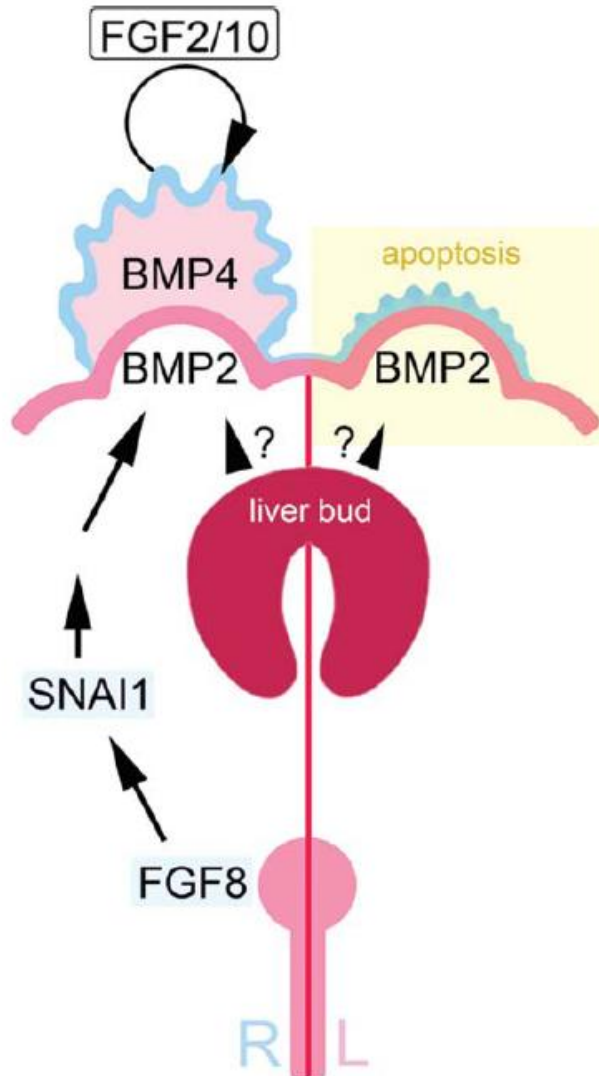
FGF promotes proliferation and survival



Reconstruction of proepicardial TBX18 expression domains after inhibition of FGF signals in vivo



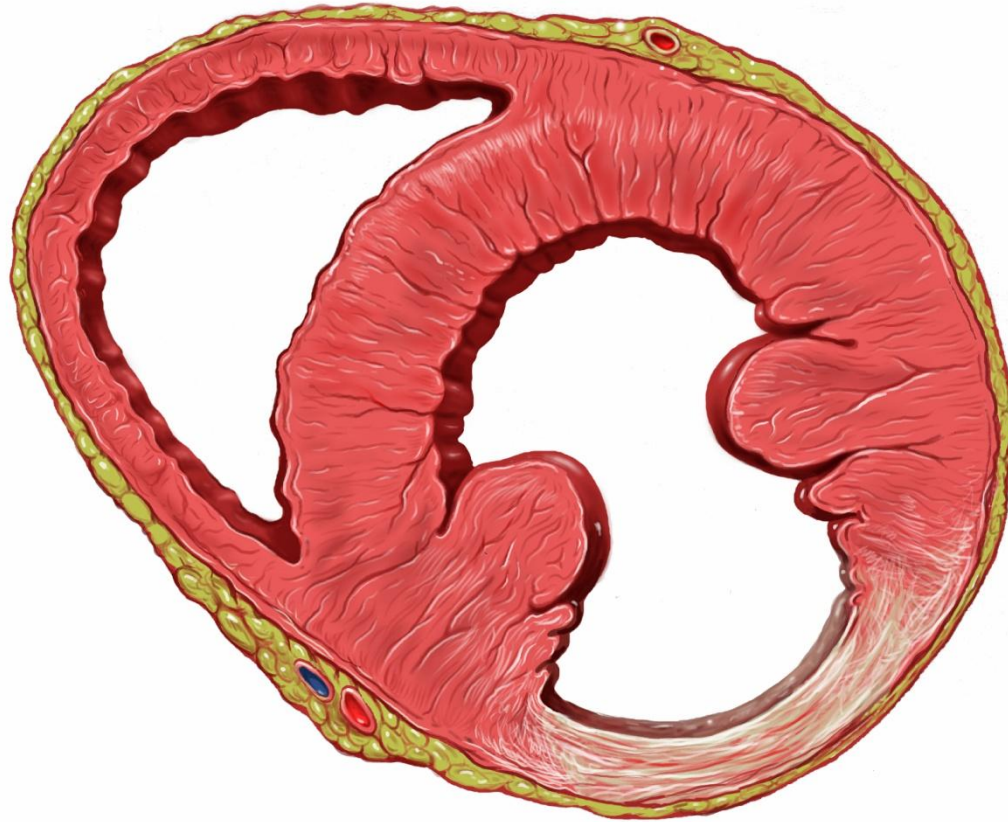
BMP and FGF signals during PE development



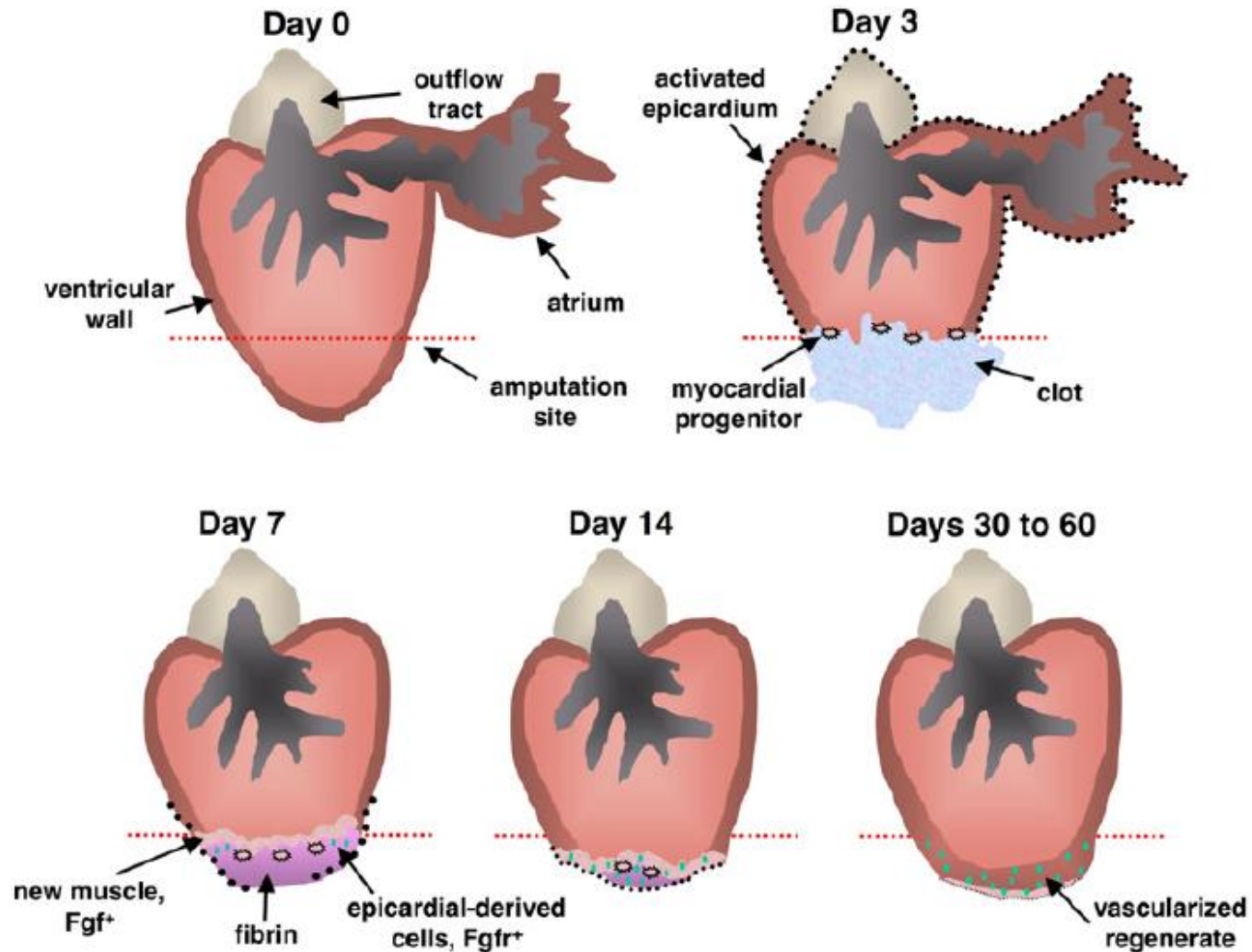
Summary 3

- BMP signalling promotes myocardial differentiation of a subset of PE cells
- High BMP dose > myocardium
- Low BMP dose > proepicardium
- FGF signalling is important for the proliferation and survival of PE cells

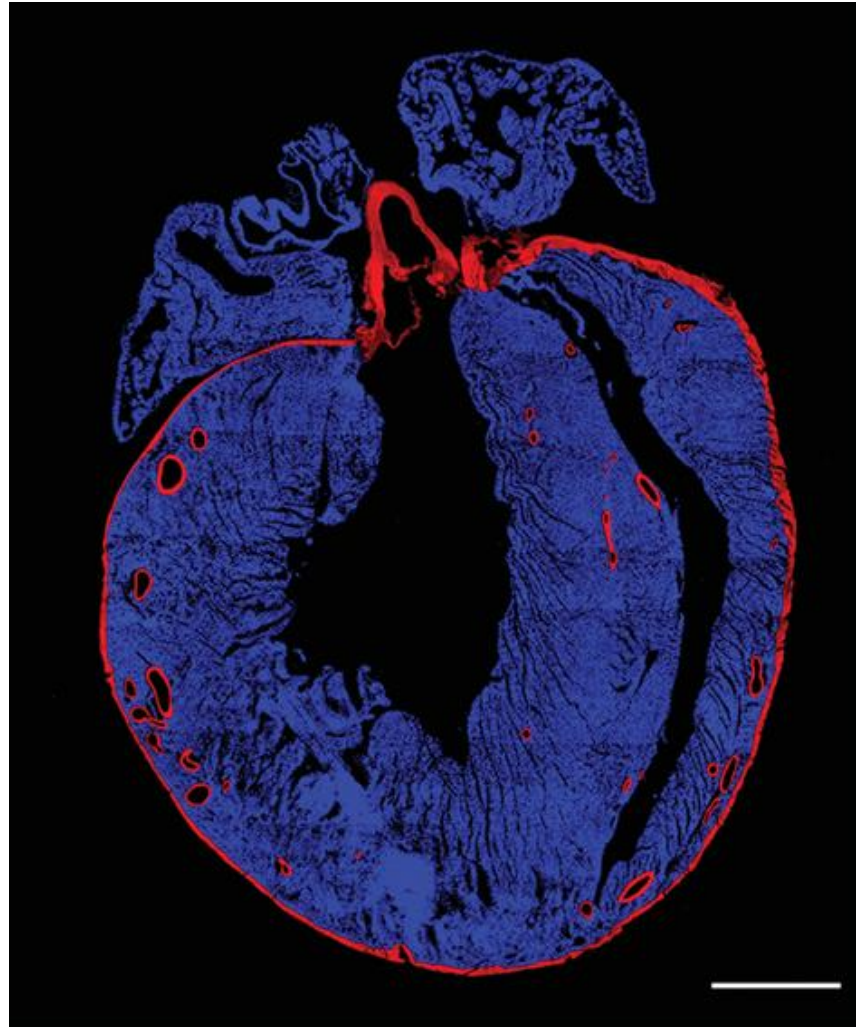
4. Neovascularization and regeneration



Process of myocardial and epicardial regeneration



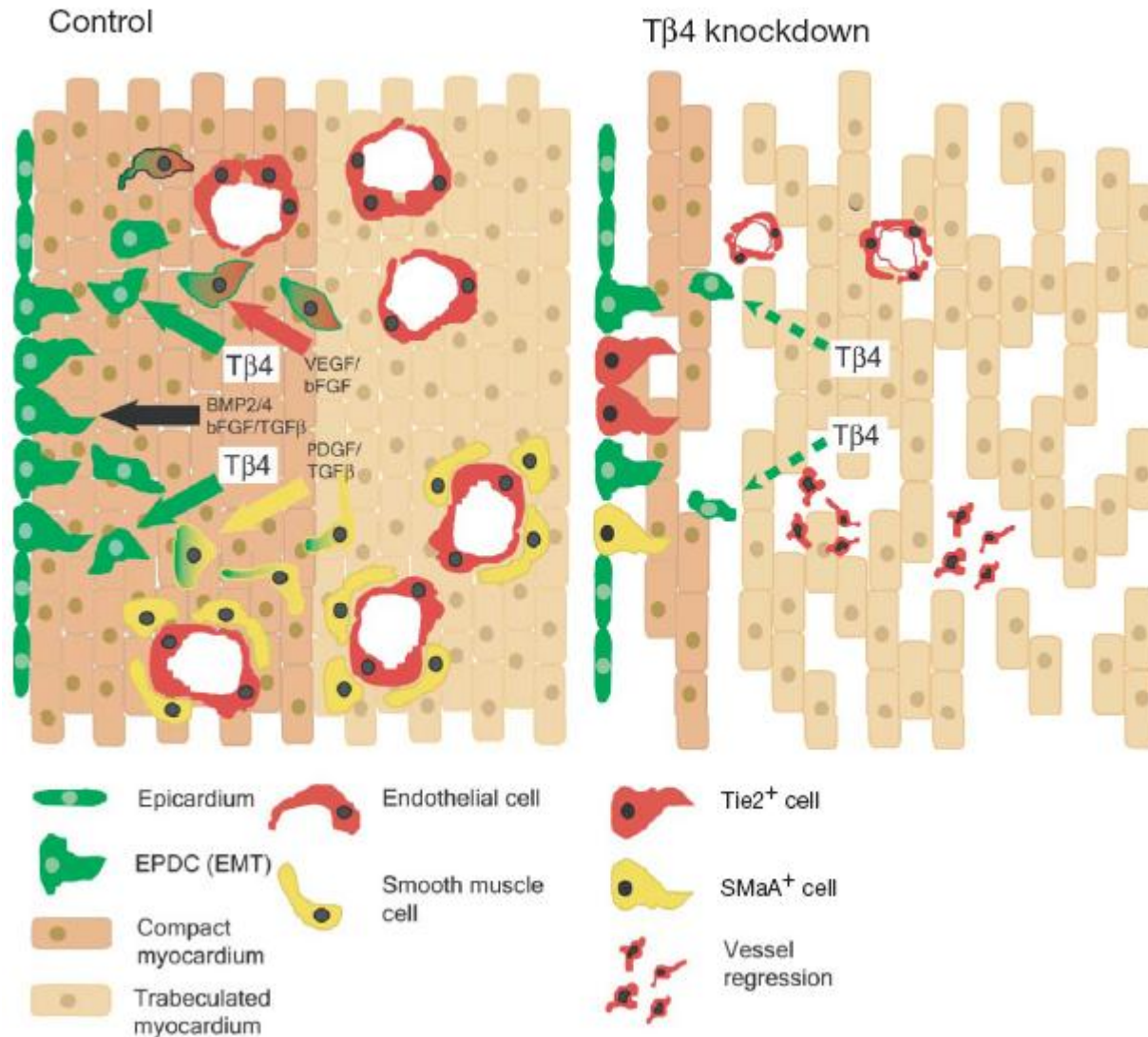
Thymosin β 4 is an important regulator for neovascularization in the mouse



Thymosin β 4

- Actin-monomer-binding protein which regulates cytoskeletal dynamics and directed cell movement
- Essential for migration of EPDCs (epicardial derived cells)
- Overexpression of T β 4 causes EPDCs to revert to their embryonic phenotype and give rise to endothelial and smooth muscle cells
- Thereby stimulating neovascularization of the heart

The function of Thymosin β 4



Summary 4

- Cardiac failure is strongly connected to ischaemic damage caused by vascular insufficiency
- The zebrafish heart is exceptional for its natural capacity to regenerate muscle tissue facilitated by reactivation of coronary vascularization
- Therefore the investigation of early developmental regulation of epicardium formation provides insight into myocardial and epicardial de/redifferentiation processes in the adult heart
- Thymosin β 4 has been shown to act as a powerful factor to regulate coronary vessels development as well as to promote neovascularization in the adult mouse heart

literature

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