

**Imperial College  
London**

BSc in Reproductive & Developmental Sciences

**Cell Biology of Preimplantation  
Development**

Kate Hardy  
Institute of Reproductive and Developmental Biology

---

---

---

---

---

---

---

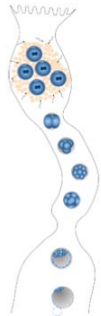
**Preimplantation development  
(mouse)**

Embryo

- Unattached
- Moves from site of fertilization (ampulla) to site of implantation (uterus)
- Depends on maternal reproductive tract for nutrition
- Undergoes cell proliferation

BUT

- No cell growth



© Kate Hardy Imperial College London

---

---

---

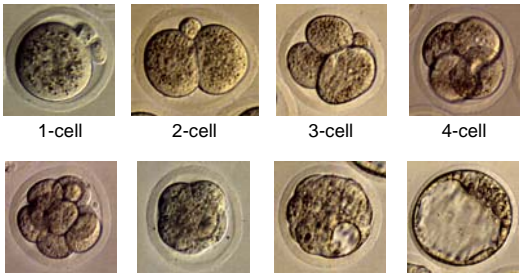
---

---

---

---

**Preimplantation development**



1-cell    2-cell    3-cell    4-cell

8-cell    morula    cavitating morula    blastocyst

© Kate Hardy Imperial College London

---

---

---

---

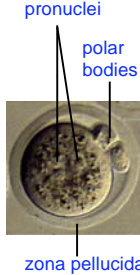
---

---

---

### Zona pellucida

- Translucent **glycoprotein** coat, 7µm thick
  - 3 glycoproteins: ZP1, ZP2 and ZP3 (mouse)
  - synthesized by oocyte during oogenesis
- **ZP3**: Primary sperm receptor
  - binds sperm head
  - activates sperm – induces acrosome reaction
  - highly conserved – 60% homology between mouse & human
  - species specific (carbohydrate side-chains)
- **ZP2**: Secondary sperm receptor
  - binds acrosome reacted sperm



© Kate Hardy Imperial College London

---

---

---

---

---

---

---

---

### Functions of zona pellucida at fertilization

- Secreted by oocyte during early folliculogenesis
- Provides **species specificity** at fertilization
  - oligosaccharides on ZP3 provide specificity
  - sperm can fuse with zona-free oocytes from other species
- Induces sperm **acrosome reaction**
- Site of '**slow block**' to polyspermy
  - cortical granule release → zona reaction
  - zona hardening
  - loss of sperm binding capacity

© Kate Hardy Imperial College London

---

---

---

---

---

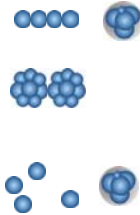
---

---

---

### Protective functions of zona pellucida

- **Maintains normal cleavage pattern**
- **Prevents sticking** of embryos to one another or to oviduct
- **Protects** embryos moving down oviduct
  - Zona-free cleavage stage embryos disintegrate in oviduct
  - Protects against attack by leukocyte



© Kate Hardy Imperial College London

---

---

---

---

---

---

---

---

### Oocyte: entry into meiosis and arrest

**PROPHASE**

- occurs during fetal life
- homologous chromosomes pair (1 from mother, 1 from father)
- chiasma form
- crossing over occurs
- oocyte arrested in primordial follicle for up to 50 years

**RESUMPTION OF MEIOSIS**

- only occurs after puberty

© Kate Hardy Imperial College London

---

---

---

---

---

---

---

---

### Resumption of meiosis after LH surge

**METAPHASE I**

- nuclear membrane breaks down
- chromosomes align on equator, stabilized by chiasmata
- no centromere division
- no chromatid separation

**ANAPHASE/TELOPHASE**

- 1 chromosome goes to one pole, other goes to other
- maternal and paternal chromosomes randomly migrate to either pole
- first polar body extruded

© Kate Hardy Imperial College London

---

---

---

---

---

---

---

---

### Second meiotic division

**METAPHASE II**

- oocyte ovulated while arrested at Metaphase II

**ANAPHASE II/TELOPHASE**

- only occurs if fertilization or activation occurs
- centromeres divide
- chromatids separate
- one set of chromatids expelled in second polar body

© Kate Hardy Imperial College London

---

---

---

---

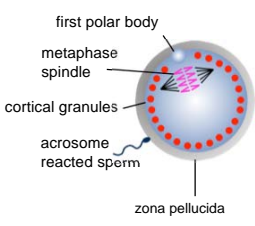
---

---

---

---

### Ovulation and fertilization



- Ovulated oocyte arrested at the second metaphase stage of meiosis
- Still surrounded by cumulus cells

© Kate Hardy Imperial College London

---

---

---

---

---

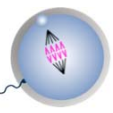
---

---

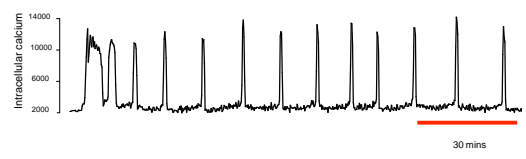
---

### Fertilization

#### Sperm fusion



- Activates calcium wave & spikes
- Induces cortical granule release, 'hardens' zona, to prevent polyspermy
- Metaphase spindle rotates
- Meiosis resumes



© Kate Hardy Imperial College London

---

---

---

---

---


---

---

---

### Fertilization (continued)

#### Pronucleus formation



- Meiotic division completes
- Second polar body extruded
- Sperm nucleus decondenses & male pronucleus forms
- Female pronucleus forms

© Kate Hardy Imperial College London

---

---

---

---

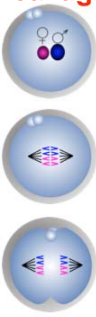
---

---

---

---

**First cleavage**



**Diploid zygote**

- pronuclei do not fuse
- chromosomes duplicate DNA
- pronuclei migrate together

**First mitotic metaphase**

- pronuclear membranes break down
- first mitotic spindle forms
- maternal and paternal chromosomes align on single metaphase plate

**First anaphase**

- chromatids separate and move apart
- cleavage furrow forms

1 chromosome of each pair from mother  
1 chromosome of each pair from father

© Kate Hardy Imperial College London

---

---

---

---

---

---

---

---

**Timing of first cell cycle (mouse)**

• Extrusion of second polar body	2 – 5 h
• Formation of male pronucleus	4 – 7 h
• Formation of female pronucleus	6 – 9 h
• DNA replication	11 – 18 h
• Cleavage to 2-cell	17 – 20 h

© Kate Hardy Imperial College London

---

---

---

---

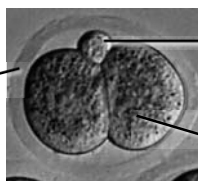
---

---

---

---

**2-cell embryo**



zona pellucida

polar body

blastomere

© Kate Hardy Imperial College London

---

---

---

---

---

---

---

---

**Meanwhile – what is happening to gene expression?**

**TRANSCRIPTION**

**TRANSLATION**  
occurs on ribosomes (rRNA & proteins)  
Amino acids added on to growing polypeptide chain by tRNA

DNA  
↓  
mRNA  
↓  
Protein

© Kate Hardy Imperial College London

---

---

---

---

---

---

---

---

**Parental → embryonic transition**

highly differentiated gametes

NO new transcription

totipotent embryonic cell

© Kate Hardy Imperial College London

---

---

---

---

---

---

---

---

**In absence of transcription**

- Molecular changes, necessary to accomplish transition from differentiated gamete to undifferentiated embryonic cell, rely on:
  - stability and differential translation of maternal mRNAs
  - stability of proteins
  - post-translational processing

© Kate Hardy Imperial College London

---

---

---

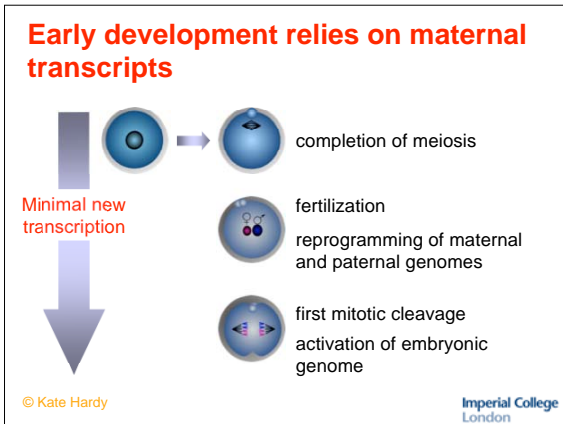
---

---

---

---

---




---

---

---

---

---

---

---

---

### Maternal transcripts accumulated during oocyte growth and maturation

- Mouse oocytes grow from 15 - 80  $\mu\text{m}$  diameter
- Human oocytes grow 35 - 120  $\mu\text{m}$  diameter
- Fully grown mouse egg contains 25 $\mu\text{g}$  protein and 0.3 - 0.55 ng RNA
- Growing oocytes contain large nucleoli and synthesize large amounts of RNA

© Kate Hardy Imperial College London

---

---

---

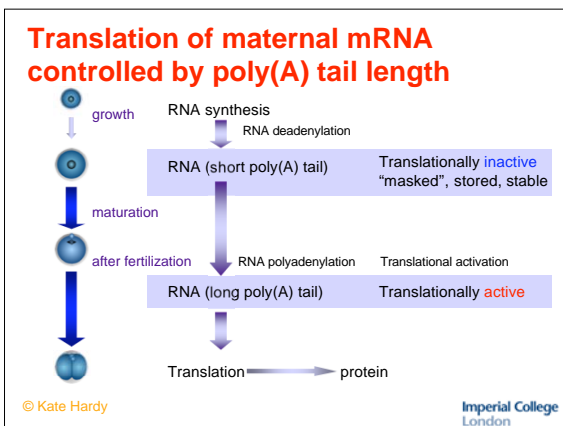
---

---

---

---

---




---

---

---

---

---

---

---

---

**Formation of pronuclei**

<b>Male</b>	<b>Female</b>
<ul style="list-style-type: none"><li>• Breakdown of nonpermeable sperm nuclear envelope</li><li>• Reformation of permeable pronucleus nuclear membrane</li><li>• Decondensation<ul style="list-style-type: none"><li>• removal of protamines</li><li>• replacement with histones</li></ul></li></ul>	<ul style="list-style-type: none"><li>• Completion of meiosis</li><li>• Formation of female pronucleus</li><li>• <b>Pronuclei remain separate until first mitosis</b></li></ul>

© Kate Hardy Imperial College London

---

---

---

---

---

---

---

---

**Nuclear changes during cleavage**

- Histone composition continues to change
- Acetylation of histones changes, facilitating transcription
- Embryo generally acquires **transcriptionally repressive** chromatin state: for tightly regulated gene regulation
- Specific genes can then be transcribed in a regulated manner (enhancers, transcription factors etc)

© Kate Hardy Imperial College London

---

---

---

---

---

---

---

---

**Activation of the embryonic genome**

Transition from maternal control to embryonic genome control marked by:

- **Degradation of maternal mRNA**
- **Activation of transcription** of embryonic genome, with transcription of new mRNAs not seen in oocyte including gene products which influence:
  - metabolism,
  - rate of cell proliferation,
  - embryo cell number &
  - differentiation.

© Kate Hardy Imperial College London

---

---

---

---

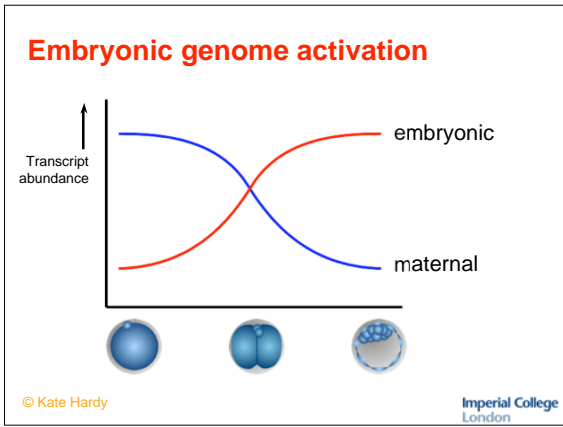
---

---

---

---





---

---

---

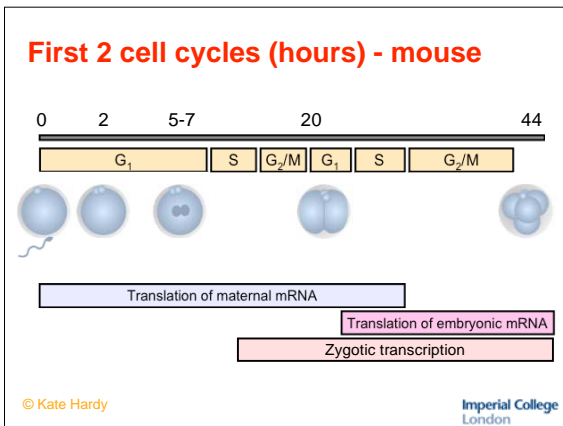
---

---

---

---

---



---

---

---

---

---

---

---

---

### Cleavage

- asynchronous cell division
- no growth
- at each division, cell volume halves

© Kate Hardy Imperial College London

---

---

---

---

---

---

---

---

### Cleavage



- cells spherical
- loosely attached
- totipotent

© Kate Hardy Imperial College London

---

---

---

---

---

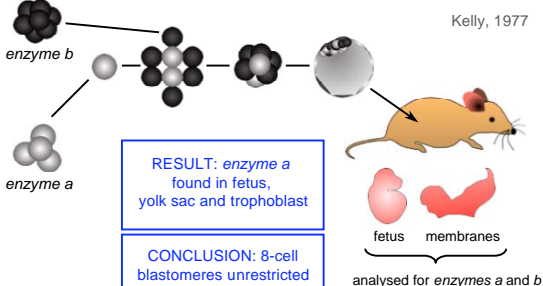
---

---

---

### Totipotency during early cleavage

Kelly, 1977



RESULT: enzyme *a* found in fetus, yolk sac and trophoblast

CONCLUSION: 8-cell blastomeres unrestricted in developmental potential

analysed for enzymes *a* and *b*

© Kate Hardy Imperial College London

---

---

---

---

---


---

---

---

### Compaction and morula formation

- Occurs at 8-cell stage (mouse), 16-cell stage (human)
- Mediated by **E-cadherin** (adhesion glycoprotein)
- Blastomeres:
  - flatten and maximize **intercellular contacts**
  - become more **adhesive**
  - become **polar** - embryo develops radial axis



© Kate Hardy Imperial College London

---

---

---

---

---

---

---

---

### Morula (mulberry)



© Kate Hardy

Imperial College London

---

---

---

---

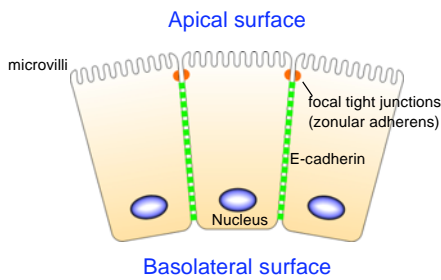
---

---

---

---

### Polarization



© Kate Hardy

Imperial College London

---

---

---

---

---

---

---

---

### E-cadherin (uvomorulin)

- Involved in **establishment** and **maintenance** of epithelial morphology
- Expressed in egg (cytoplasm) and embryo (cell surface)
- Redistributed to cell contact points at 8-cell stage
- Responsible for  $Ca^{2+}$ -dependent compaction
  - 8-cell morulae decompact in  $Ca^{2+}$ -free medium
  - antibodies to E-cadherin prevent compaction
- Maybe mediated by protein phosphorylation (8c)
- Mice lacking E-cadherin
  - compact (residual maternal transcripts) **but**
  - fail to form trophectoderm or a blastocoel cavity

© Kate Hardy

Imperial College London

---

---

---

---

---


---

---

---

### Morula: formation of 2 cell types

- Polarization of cells at compaction is critical for formation of 2 different cell types in blastocyst
  - outer polar **trophoblast** (TE)
  - inner apolar **inner cell mass** (ICM)
- **Differential adhesiveness** maintains this pattern
- Morula formation marks **first differentiation** in embryogenesis



© Kate Hardy Imperial College London

---

---

---

---

---

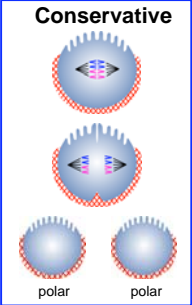
---

---

---

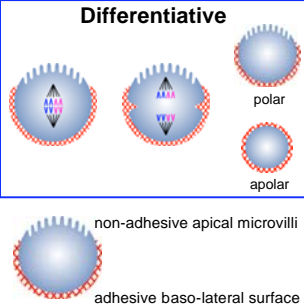
### Morula: formation of 2 cell types

#### Conservative



polar polar

#### Differentiative



polar  
apolar

non-adhesive apical microvilli  
adhesive baso-lateral surface

© Kate Hardy Imperial College London

---

---

---

---

---

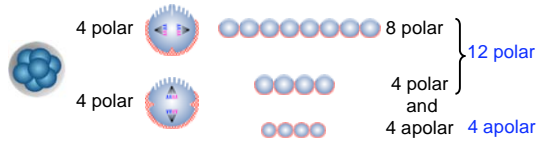
---

---

---

### Allocation to TE and ICM

8-cell → 16-cell



4 polar 8 polar } 12 polar  
4 polar and 4 apolar } 4 apolar

© Kate Hardy Imperial College London

---

---

---

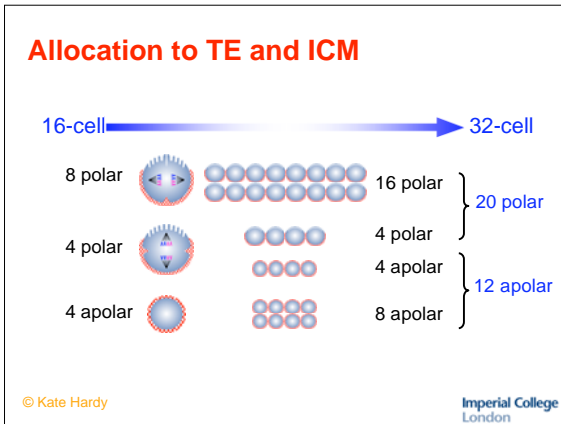
---

---

---

---

---




---

---

---

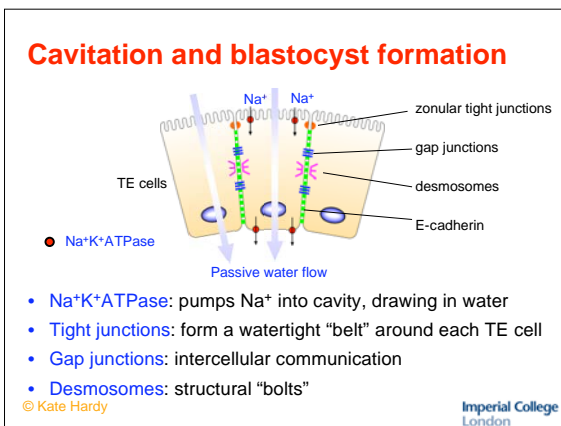
---

---

---

---

---




---

---

---

---

---

---

---

---

- ### Cavitation and blastocyst formation
- Formation of fluid filled cavity (function unknown)
    - Occurs at completion of fifth cleavage division (mouse: 16- to 32-cell stage)
  - Cell number at cavitation varies with species
    - mouse 32-cell
    - human 32-cell
    - rabbit 128-cell
    - pig 16-cell
    - sheep 64-cell
  - Independent of
    - cell number (half and double embryos cavitate at same time)
    - number of cell divisions
    - DNA replication
- © Kate Hardy Imperial College London

---

---

---

---

---

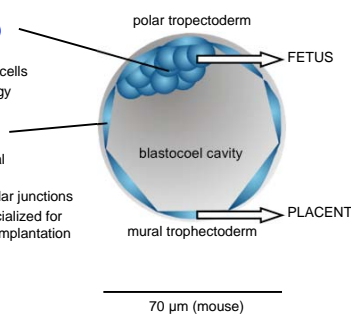
---

---

---

### Blastocyst structure

- **Inner cell mass (ICM)**
  - apolar
  - uniformly adhesive cells
  - stem cell morphology
  - pluripotent
- **Trophectoderm (TE)**
  - polar cells, epithelial morphology
  - extensive intercellular junctions
  - differentiated - specialized for fluid transport and implantation



70 μm (mouse)

© Kate Hardy Imperial College London

---

---

---

---

---

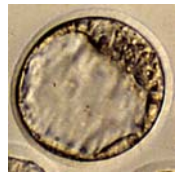
---

---

---

### Functions of trophoblast

- pump fluid into cavity
- transport metabolites between maternal tissues and ICM
- initiate chemical and physical communication between fetus and mother
- provide a proliferative source of cells for placental TE



© Kate Hardy Imperial College London

---

---

---

---


---

---

---

---

### Blastocyst expansion and hatching



- **Expansion**
  - enlargement of blastocoel cavity, expansion of blastocyst
  - continued cell division, especially in TE
  - cell number in ICM remains at ~15 - 20, until implantation
  - cell death, especially in ICM
  - ICM differentiates
- **Herniation and hatching**
- **Attachment and implantation**

© Kate Hardy Imperial College London

---

---

---

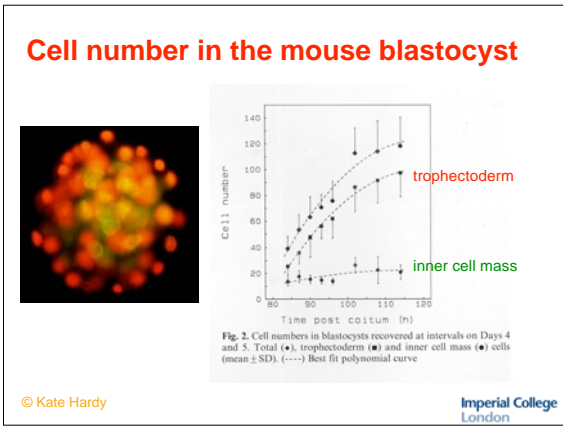
---

---

---

---

---




---

---

---

---

---

---

---

---

### Apoptosis in the mouse blastocyst

- Apoptosis occurs in ICM
- Role unknown
  - Remove cells with potential to form TE?
  - Remove defective cells?
  - Maintain ICM size?
- Signalling pathways unknown
  - BCL-2 family members and caspase genes present

© Kate Hardy Imperial College London

---

---

---

---

---

---

---

---




---

---

---

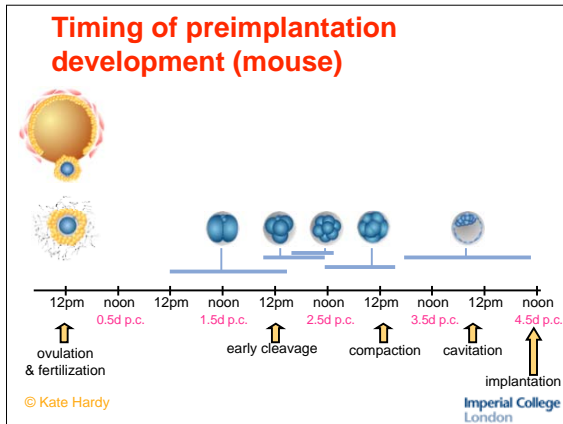
---

---

---

---

---




---

---

---

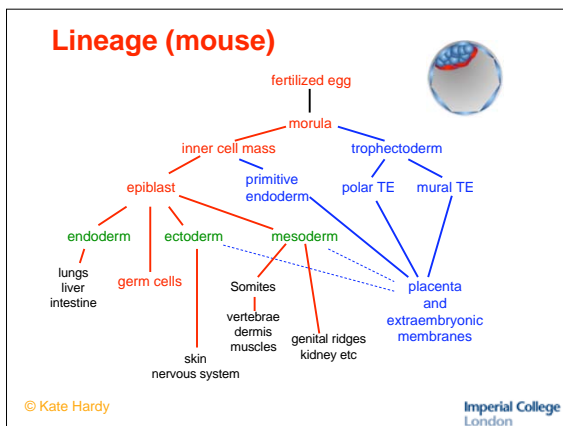
---

---

---

---

---




---

---

---

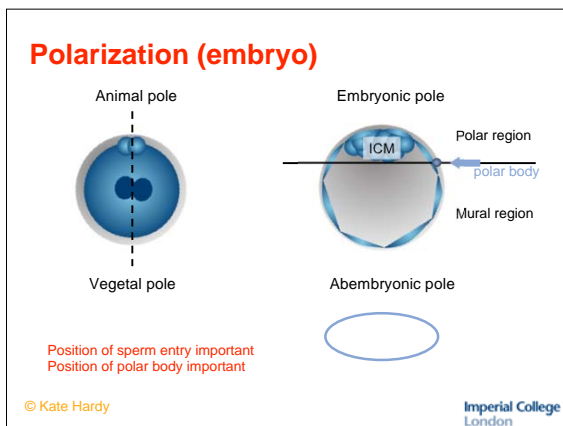
---

---

---

---

---




---

---

---

---

---

---

---

---



**Comparative preimplantation development**

- All mammalian species similar overall
- Differences in
  - Size (small)
  - Time in oviduct
  - Length of cell cycles
  - Timing of switch-on of embryonic genome
  - Timing of compaction/cavitation
  - Cell number at compaction/cavitation
  - Timing of implantation
  - Cell number at implantation

© Kate Hardy

Imperial College  
London

---

---

---

---

---

---

---

---