#### **BSc Pharmacology and Translational Medicine**

Module: Principles of Pharmacodynamics and Pharmacokinetics

Lecture: Structural Analysis of Drugs and Their Targets

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# **Structural Analysis of Drugs and Their Targets**

#### Lecture Outline

- Structural genomics
- Structural analysis
  - X-ray crystallography
  - NMR spectroscopy
    - Macromolecules
    - Small molecules
    - Interaction experiments
  - Mass Spectrometry
    - LC/MS
- Summary

#### **Structural Analysis - Outline**

•Knowledge of the chemical structure of biological molecules, and how these molecules are arranged in three dimensions can help us understand molecular interactions

•Understanding drug-target interactions are vital in pharmacology:

- To explain the observed effects of different agents
- In the development of novel therapies that act on a particular molecular target

•The majority of pharmacological interactions are either protein-protein or ligand-protein

•Spectroscopic tools to characterise proteins and small molecules are available and the most commonly used as they provide high-resolution, information rich data about molecular conformations and interactions.

### **Structural Genomics**

#### **Structural Analysis – Structural Genomics**

 Structural genomics is the area of research concerned with solving the 3D structures of coding DNA

•Eventually will provide a library of structures that can be used in a complementary fashion, alongside genomics and other information

Existing solved structures can help speed up future efforts due to the conserved nature of many macromolecular domains (e.g. binding sites in proteins). To date, >44000 macromolecular structures have been solved

•Several techniques are available for characterising the 3D conformation of macromolecules

•Two techniques can provide atomic resolution structures

X-Ray Crystallography

NMR Spectroscopy

•Other techniques (e.g. mass spectrometry) can assist in characterising a variety of molecules

### X-Ray Crystallography

#### X-Ray Crystallography Selection of target protein A method of determining the crystal structure of a compound Cloning and expression of the recombinant protein The scattering/diffraction pattern observed when X-rays are applied to Solubility and stability tests: the compound can allow the spatial location of the atom centres in the optimization of protein expression molecule to be calculated Large-scale purification Requires a considerable quantity compound as a crystal and success of the X-ray diffraction experiment is largely reliant on the **quality/purity** of Crystallization screening the crystal Crystal optimization Crystallisation conditions for a particular protein will vary and usually a large number of conditions must be tried before a pure crystal can be Data collection and structure determination obtained Functional inferences. A large number of different approaches including: comparison with similar structures, establishment of biochemical pathways Vapor diffusion Dialysis Potential drug development Microfluidics See Chayen E. and Saridakis et al. 2008

#### X-Rays on the Electromagnetic Spectrum

Electronic transitions: X-rays / Ultra-violet (UV)

Vibrational transitions: Infra-red (IR)

Nuclear spin transitions: Radio waves (NMR)



#### X-Ray Diffraction

- Protein crystals are bombarded with high energy X-rays
- A small proportion of the beam is refracted/scattered by the atoms in the crystal
- In a well-ordered (pure) crystal, a clear diffraction pattern can be observed using a detector
- A 3D electron density map can then be produced
- •Knowledge of the amino acid sequence then allows the 3D structure to be calculated
- •The resolution of the X-ray diffraction data determines how similar the model fit will be to reality



#### X-Ray Crystallography

Benefits:

Good quality crystals can lead to high-resolution diffraction patterns

•Very efficient structure solving using established algorithms

Robotics / high-throughput technologies can be employed

- Rapid identification of optimal crystal growth conditions
- Automation of main steps efficient
- Generation of large quantities of protein for analysis

#### X-Ray Crystallography

•Limitations:

•The resolution of the diffraction patterns observed in X-ray crystallography experiments is partly determined by the size of the molecule in the crystal

Larger molecules cannot be very well resolved (e.g. large proteins)

•Some proteins are difficult to crystallise

Affected by shocks and temperature fluctations

Some molecules crystallise in multiple orientations, distorting the resulting diffraction pattern

Exposure to X-rays may cause the protein to denature/degrade

A destructive process

Dynamics and in situ experiments are not possible

•Experiments with ligand binding etc require separate crystals to be obtained

### **NMR Spectroscopy**

#### NMR Spectroscopy – Outline

•Nuclear Magnetic Resonance (NMR) spectroscopy is one of the most powerful tools in modern science. Since its discovery >50 years ago, it has spread from physics to chemistry, biological science, materials science and medical diagnosis.

Phenomenon of nuclear magnetic resonance (NMR) discovered in 1945

■Bloch *et al.* ■Purcell *et al.* 

Awarded Nobel Prize in Physics (1952) for the first experimental verifications of the phenomenon

•NMR is still the only method of determining the structure of a molecule in a liquid state (c.f. crystallography)



#### NMR Spectroscopy - Outline

Nuclear Magnetic Resonance (NMR) Spectroscopy

- •Generates spectra that describe the chemical environment of nuclei in biofluid molecules
  - Chemical structure
  - Interactions
  - Bulk properties can be studied
  - Diffusion
  - Compartmentation
- Typically concentrate on <sup>1</sup>H
- Ubiquitous in biomolecules and drugs
- Good sensitivity
- LOD of <sup>1</sup>H NMR typically in micromolar range

#### **NMR Spectroscopy – Outline**

•At thermal equilibrium in the Earth's magnetic field, the numbers of nuclear spins are almost equal between the upper and lower energy levels

Increasing the field by millions of times gives workable energy differences



#### NMR Spectroscopy - Outline

When placed in a magnetic field there will be a small net difference in the populations orient
Boltzmann distribution - Net magnetisation in direction of field



•By **disturbing the equilibrium condition** and observing the resulting **return over time**, we can determine the **resonance frequencies** of the nuclei in a sample

Frequency is related to chemical environment

When scaled to the magnet strength = chemical shift

#### NMR Spectroscopy on the Electromagnetic Spectrum

Electronic transitions: X-rays / Ultra-violet (UV)

- Vibrational transitions: Infra-red (IR)
- Nuclear spin transitions: Radio waves (NMR)





#### NMR Spectroscopy

Nobel Prize in Chemistry (1991)

■R.R. Ernst

Nobel Prize in development of NMR techniques

- In particular Fourier Transform (FT)-NMR
- Fourier transformation (FT)

•Mathematical transformation of time dependent signal into frequency domain











#### NMR Spectroscopy – <sup>13</sup>C NMR Chemical Shifts



#### •NMR Spectroscopy - <sup>1</sup>H NMR Spectrum of Ethanol



•NMR Spectroscopy - <sup>1</sup>H NMR of compound from suspected drugs haul



#### NMR Spectroscopy

•K. Wüthrich shared the Nobel Prize in Chemistry (2002) for his work on the structure elucidation of large biomolecules such as proteins & nucleic acids







#### NMR Spectroscopy - Two dimensional simplification (HSQC spectrum)





#### NMR Spectroscopy - Example

Human PTB protein

Target of viral hijacking

- Structure can be solved by NMR
- •PTB = polypyrimidine tract binding protein



### NMR Spectroscopy – Overlay of 2D Spectra

No RNA RNA Added



# NMR Spectroscopy – Overlay of 2D Spectra (Zoomed)No RNARNA Added



#### **NMR Spectroscopy - Interactions**

 Projection of frequency change onto the protein structure can reveal the site of interaction

Large change in NMR frequency

No change in NMR frequency



#### **NMR Spectroscopy - Summary**

 Molecular modelling approaches can be used to visualise how molecules interact



NMR Spectroscopy – Information on Spatial Proximity

2D-Nuclear Overhauser Effect (NOE) experiments

Couplings across space – stereo structure (peptides, proteins, etc.)



#### NMR Spectroscopy - Summary

- ■Some nuclei (e.g. <sup>1</sup>H, <sup>13</sup>C) are accessible to NMR spectroscopy
- •NMR spectroscopy observes effect of disturbing spin populations using electromagnetic radiation
- •Frequency, intensity and fine structure are the three principal features of an NMR spectrum
- Can be applied to structure elucidation, quantification, and medical diagnosis/imaging
- Powerful tool in metabonomics for profile generation
- Easily translatable results
- High resolution NMR MAS of intact tissue MRI in vivo

### **Mass Spectrometry**



#### **MS** - Outline

Characterises components by:
Mass-to-charge ratio (*m/z*)
Fragmentation pattern

- Used for structure elucidation
  - Masses of fragments give partial structural information
  - Tryptic digest can also be used information on resulting peptides acquired

A variety of spectrometers are available

 Q-ToF and FTMS provide very high *m/z* accuracy/precision that can allow deduction of the empirical formula

Note: other MS-based techniques are routinely used in structural analysis

- MALDI/MS
- GC/MS
- •CE/MS..... etc



#### LC/MS - Summary

- MS can be used with chromatography
  - Separates components
  - Helps deconvolve complex mixtures
  - •Retention times are usually very stable and provide an additional characteristic for i.d.
- •Very sensitive (pmol) technique to detect minor components
- •Used extensively in combination with NMR for rapid structure elucidation of unknowns (e.g.)



LC/MS - Setup





#### LC/MS – Electrospray Ionisation

- Atmospheric pressure ionisation
- Soft ionisation technique





#### LC/MS – Separation and Detection

#### ■3D data

- ■Mass-to-charge ratio (*m*/*z*)
- Time
- Intensity



### LC/MS

#### LC/MS – Plasma Chromatogram





#### LC/MS – Spectrum





#### LC/MS

Benefits:

- Rapid method for structural information
- Separates biofluid components and used in metabolism / PK studies
- Easily automated
- Information-rich datasets



#### LC/MS

•Limitations:

- Infusion MS generates very complex, overlapped spectra
- •LC/MS is experimentally more complicated and generates (typically) very large datasets

Mass spectra of trypic digests of proteins are very complex:Require substantial expertise and computational power

•Suffers from ion suppression:

Can make observation/quantification of affected compounds difficult

### **Lecture Summary**

#### **Main Points**

•Elucidating the structure of drugs and their targets is important in understanding the molecular mechanisms that they employ and can lead to the targeted development of new therapies

The two main methods for characterising the 3D structure of large molecules such as proteins are:
X-ray crystallography (calculation of structure using diffraction pattern of a crystal)
NMR spectroscopy (calculation of structure using connectivity and distance measures)

•NMR is also used extensively to charaterise small molecules and metabolites, especially in complex mixtures

 Additional information can be obtained using complementary techniques such as mass spectrometry

Characterising the interaction of drugs and their targets is possible using these and other techniques.